

## Supporting Information

## Synthesis and Resolution of a Novel Chiral Diamine Ligand and Application to Asymmetric Lithiation-Substitution

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**General.** Unless otherwise stated, all non-aqueous reactions and distillations were carried out under an atmosphere of dry nitrogen in glassware that had been either flame-dried under a stream of nitrogen or in an oven (180 °C) for at least 12 h. When necessary, solvents and reagents were dried prior to use. Toluene, THF, Et<sub>2</sub>O, and CH<sub>2</sub>Cl<sub>2</sub> were de-oxygenated by purging with Ar and then dried by passing through activated alumina. MeOH was distilled from Mg(OEt)<sub>2</sub>. Benzene, n-pentane, MTBE and TMEDA were distilled from Na and MeCN from CaH<sub>2</sub>.

A 450 mL reactor (No. N4767 from the Parr Company) was used for the high-pressure reactions. Reactions were stirred by an external magnetic stir plate and heated with an external high-temperature silica oil bath. Reported temperatures are of the external bath.

Analytical thin layer chromatography (TLC) was performed on EM Reagents 0.25 mm silica gel 60-F plates. Visualization was accomplished with UV light. Chromatography on silica gel was performed using a forced flow of the indicated solvent system on EM Reagents Silica Gel 60 (230-400 mesh). HPLC analyses were performed using a Waters Delta 600 system ( $\lambda = 254$  nm) connected to Chiralpak AD column (4.6 x 150 mm) from Daicel. GC analyses were performed on Hewlett-Packard 5890A system with a Cyclodex-B-Fused Silica Capillary column (30 m x 0.25 mm) from Alltech. Melting points were obtained on Thomas Scientific Unimelt apparatus and are uncorrected.

<sup>1</sup>H NMR spectra were recorded on Bruker AM-500 (500 MHz), AM-250 (250 MHz), or AM-200 (200 MHz) spectrometers. <sup>13</sup>C NMR spectra were recorded on a Bruker AM-500 (125 MHz) spectrometer. Chemical shifts are reported in ppm from tetramethylsilane (0 ppm) or with the solvent resonance as the internal standard (CDCl<sub>3</sub> 7.26 ppm, DMSO-d<sub>6</sub> 2.49 ppm, D<sub>2</sub>O 4.80 ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, br = broad, m = multiplet), coupling constants, and number of protons. Mass spectra were obtained on a low resonance Micromass Platform LC in electron spray mode. IR spectra were taken on a Perkin-Elmer FT-IR spectrometer using thin films or in a cell using CHCl<sub>3</sub> solution with CHCl<sub>3</sub> as background.

**cis-Decahydro-1,5-naphthyridine (10).** To 1,5-naphthyridine (8.15 g, 62.7 mmol) dissolved in AcOH was added 5% Rh/Al<sub>2</sub>O<sub>3</sub> (0.817 g) and acetic acid (70 mL). The mixture was hydrogenated at 100 °C and 1200 psi H<sub>2</sub> for 12 hours. After filtration, the residue was washed with MeOH saturated with HCl. Removal of the solvent gave a solid (10.5 g, 95%) which was recrystallized from MeOH and H<sub>2</sub>O to yield the *trans* HCl salt. At this point the mother liquor was an 85:15 mixture of *cis*-**10** and the *trans* isomer in the form of HCl salt. The solvent was removed from this mixture and the salt was dissolved in H<sub>2</sub>O. The aqueous solution was made basic with NaOH, saturated with NaCl, and extracted with Et<sub>2</sub>O. The combined Et<sub>2</sub>O extracts were dried over anhydrous K<sub>2</sub>CO<sub>3</sub>. Removal of Et<sub>2</sub>O afforded a 94:6 mixture of *cis*-**10** and the *trans* isomer as the free diamine. Vacuum distillation (89–90 °C, 3 mm Hg) afforded pure **10** (4.2 g, 48%) as a waxy white solid: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.26–1.40 (m, 2H), 1.51–1.68 (m, 8H), 2.56–2.64 (dt, J = 3.0, 12.0 Hz, 2H), 2.68 (s, 2H) 3.04–3.11 (dd, J = 3.0, 12.0 Hz, 2H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  21.2, 31.0, 47.0, 53.6; IR (CHCl<sub>3</sub>) 3017, 2937, 1206 cm<sup>-1</sup>; MS (ESI, MH<sup>+</sup>, C<sub>8</sub>H<sub>17</sub>N<sub>2</sub>) 141.

**(*R,R*)-cis-Decahydro-1,5-naphthyridine (10) (*R,R*)-Tartrate.** Racemic **10** (0.98 g, 7.0 mmol, *cis:trans* 94:6) was dissolved in MeOH (5 mL) and added slowly to a MeOH (5 mL) solution of L-(+)-tartaric acid (525 mg, 3.5 mmol) at rt. After the solution clarified, acetic acid (0.36 mL, 6.3 mmol) was added. A few drops of Et<sub>2</sub>O were added to precipitate out a white powder. H<sub>2</sub>O (2 mL) was added and the mixture was warmed to dissolve the precipitate. Upon standing at 0 °C overnight the tartrate salt of **10** (887 mg, 44%) was isolated in 87% ee. Recrystallization of a portion (550 mg) from EtOH (4 mL) and H<sub>2</sub>O (1 mL) caused enrichment to ≥98% ee (458 mg, 37% overall out of a 50% theoretical yield). The mother liquors could be combined, the solvent removed, and then treated in a similar manner with D-(-)-tartaric acid to yield (*S,S*)-**10** (*S,S*)-tartrate: (*R,R*)-**10** (*R,R*)-tartrate >99% ee [ $\alpha$ ]<sub>D</sub><sup>25</sup> = +24.5 (c 2.96, H<sub>2</sub>O); (*S,S*)-**10** (*S,S*)-tartrate >99% ee [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -25.3 (c 2.96, H<sub>2</sub>O); <sup>1</sup>H NMR (500 MHz, D<sub>2</sub>O)  $\delta$  1.84-1.91(m, 2H), 2.00-2.10 (m, 4H), 2.17-2.23 (m, 2H), 3.21-3.25 (m, 2H), 3.32-3.37 (m, 2H) 3.86-3.87(m, 2H), 4.36 (s, 2H); <sup>13</sup>C NMR (125 MHz, D<sub>2</sub>O)  $\delta$  19.7, 22.9, 42.3, 51.4, 75.2, 179.6.

Enantiomeric excess was determined by isolating the free amine from a small sample of the salt *via* extraction and then acylating with benzoyl chloride using DMAP and Et<sub>3</sub>N (see compound **11e** below). The resultant bisamide was easily resolved using a chiral HPLC column (Chiralpak AD: *t*<sub>R</sub> (*R,R*) = 8.5 and *t*<sub>R</sub> (*S,S*) = 22.4 min; 1.0 mL/min; 70:30 hexane:iPrOH).

An X-ray structure of the crystalline (*R,R*)-**10** (*R,R*)-tartrate was obtained and absolute configuration was assigned using the known absolute stereochemistry of L-(+)-tartaric acid (tables of X-ray data for this compound are included at the end of the supporting information).

Enantiomerically pure (*R,R*)-*cis*-decahydro-1,5-naphthyridine (**10**) was obtained by dissolving the diastereomerically pure tartrate salt in aqueous NaOH saturated with NaCl and extracting with Et<sub>2</sub>O. Drying the combined Et<sub>2</sub>O extracts over anhydrous Na<sub>2</sub>SO<sub>4</sub> and removal of Et<sub>2</sub>O afforded enantiomerically pure (*R,R*)-**10** as a white solid: (*R,R*)-**10** 98% ee [ $\alpha$ ]<sub>D</sub><sup>25</sup> = +33.5 (c 0.74, EtOH); (*S,S*)-**10** 93% ee [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -32.0 (c 0.78, EtOH)

***N,N'*-Dimethyl-*cis*-decahydro-1,5-naphthyridine (11a).** Diamine **10** (730 mg, 5.21 mmol) was mixed with formaldehyde (4 mL, 37% w/w aq solution) and formic acid (2 mL 88% w/w. aq. solution) cautiously. The mixture was kept at 70 °C in an oil bath for 24 h. Water was added and then 50% aq. NaOH was used to adjust the pH to >14. The solution was saturated with NaCl and extracted with Et<sub>2</sub>O. The combined Et<sub>2</sub>O extracts were dried over K<sub>2</sub>CO<sub>3</sub>. Filtration and concentration afforded a crude oil which was subjected to bulb-to-bulb distillation at 80 °C (0.5-1 mm Hg) to yield colorless oil **11a** (760 mg, 87% yield): (*R,R*)-**11a** ≥98% ee [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -9.9 (c 0.81, EtOH); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) 1.2-1.6 (br, 4H), 1.7-2.0 (br, 4H), 2.1-2.5 (br, 10H) 2.5-3.1 (br, 2H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  21.7 (br), 28.0 (br), 42.4, 57.5 (br), 62.1 (br); MS (ESI, MH<sup>+</sup>, C<sub>10</sub>H<sub>21</sub>N<sub>2</sub>) 169.

**Typical Two-Step Sequence for *N,N'*-Disubstituted Amines 11b-f.** Compound **10** in the form of the HCl salt or free amine (1.0 equiv), Et<sub>3</sub>N (5.0 equiv) and DMAP (0.2 equiv) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> at 0 °C. An acid chloride (4.0 equiv) was added slowly *via* syringe. The resultant mixture was stirred at rt overnight. The insoluble solid was removed by filtration. Concentration of the filtrate and purification by chromatography yielded the pure *cis*-bisamide.

Bisamide (1.0 equiv) was dissolved in anhydrous THF. LiAlH<sub>4</sub> (4.0-5.0 equiv) was added slowly accompanied by vigorous foaming. The mixture was heated to reflux for 1 h and then stirred 12 h at rt. The reaction was quenched by adding aqueous NaOH solution following the Fieser protocol. Filtration through Celite<sup>®</sup>, and removal of solvent afforded crude product which was purified by silica column chromatography or bulb-to-bulb distillation.

***N,N'*-Diethyl-*cis*-decahydro-1,5-naphthyridine (11b).** Using compound **10** and acetyl chloride, *N,N'*-diacetyl-*cis*-decahydro-1,5-naphthyridine (75%) was obtained. <sup>1</sup>H and <sup>13</sup>C NMR indicate that two rotamers present: (*R,R*)-**11b** ≥98% ee [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -7.3 (c 0.84, EtOH); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.4-1.7 (m), 1.7-1.8 (m), 1.9-2.1 (m), 2.4-2.6 (m) 2.9-3.1 (m) 3.3-3.8 (m) 4.4-4.8 (m); <sup>13</sup>C NMR (125

MHz, CDCl<sub>3</sub>) δ 21.3, 21.4, 21.7, 21.9, 22.0, 22.0, 23.1, 23.2, 24.0, 24.1, 24.9, 25.2, 35.6, 35.8, 41.0, 41.3, 47.8, 48.5, 53.6, 54.6, 169.1, 169.2, 169.3; MS (ESI, MNa<sup>+</sup>, C<sub>12</sub>H<sub>20</sub>N<sub>2</sub>O<sub>2</sub>Na) 247.4.

Reduction of the above bisamide afforded **11b** (74%) after bulb-to-bulb distillation 125 °C (1-3 mm Hg): <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.04-1.07 (t, J = 7 Hz, 6H), 1.50-1.69 (m, 8H), 2.36 (br, 2H) 2.52-2.58 (m, 6H) 3.02 (br, 2H); MS (ESI, MH<sup>+</sup>, C<sub>12</sub>H<sub>25</sub>N<sub>2</sub>) 197.4

***N,N'*-bis-Trifluoroethyl-*cis*-decahydro-1,5-naphthyridine (11c)**: Using compound **10** and trifluoroacetyl chloride, *N,N'*-trifluoroacetyl-*cis*-decahydro-1,5-naphthyridine (85%) was obtained. <sup>1</sup>H and <sup>13</sup>C NMR indicate that two rotamers present: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.57-1.91 (m), 1.93-2.04 (m), 2.12-2.25 (m), 2.80-2.87 (m), 3.11-3.18 (m), 3.84-3.86 (t, J = 14 Hz), 3.99-4.07 (m), 4.45-4.47 (m), 4.63-4.72 (m); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 21.57, 23.35, 23.52, 23.7, 23.8, 24.5, 24.6, 37.4, 37.7, 40.3, 40.3, 40.5, 40.5, 40.6, 49.5, 50.3, 52.8, 53.8, 115.2, 115.4, 117.5, 117.7, 155.8, 156.5; MS (ESI, MCl<sup>-</sup>, C<sub>12</sub>H<sub>14</sub>N<sub>2</sub>O<sub>2</sub>F<sub>6</sub>Cl) 367.2, 369.2.

Reduction of the above bisamide afforded **11c** (94%) after chromatography: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.45-1.58 (m, 4H), 1.68 (m, 4H), 2.62 (m, 4H), 2.97 (m, 4H), 3.07 (m, 2H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 17.2, 23.9, 47.0, 55.5 (J<sub>CCF</sub> = 31 Hz), 59.8, 67.9, 125.8 (J<sub>CF</sub> = 277 Hz).

***N,N'*-bis-Neopentyl-*cis*-decahydro-1,5-naphthyridine (11d)**. Using compound **10** and pivaloyl chloride, *N,N'*-trimethylacetyl-*cis*-decahydro-1,5-naphthyridine was obtained. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.1-1.4 (s, 18H), 1.4-2.0 (m, 8H), 2.75-2.95 (t, 2H) 4.0-4.2 (br, 2H) 4.4-4.6 (br, 2H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 22.3, 25.0, 27.1, 28.4, 28.5, 39.0, 176.7; MS (ESI, MNa<sup>+</sup>, C<sub>16</sub>H<sub>32</sub>N<sub>2</sub>O<sub>2</sub>Na) 331.5.

Reduction of the above bisamide afforded **11d** (81% for two steps) after chromatography: <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>) δ 0.81 (s, 18H), 1.41-1.70 (m, 8H), 1.93-2.27 (dd, J = 14 Hz, 4H), 2.29-2.36 (m, 2H), 2.51-2.74 (m, 4H); MS (ESI, MH<sup>+</sup>, C<sub>16</sub>H<sub>37</sub>N<sub>2</sub>) 281.5.

***N,N'*-Dibenzyl-*cis*-decahydro-1,5-naphthyridine (11e)**. Using compound **10** (75:25 *cis:trans* HCl salt, 1.92 g, 10.9 mmol) and benzoyl chloride (4.2 mL, 36.0 mmol), *N,N'*-dibenzoyl-*cis*-decahydro-1,5-naphthyridine (2.45 g, 65%) was obtained as a white solid after chromatography (2:1 EtOAc:hexanes): mp (±)-bisamide 168-170 °C, (*R,R*)-bisamide 198-200 °C; (*R,R*)-bisamide [α]<sub>D</sub><sup>25</sup> = -22.7 (c 0.26, EtOH); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.25-2.35 (m, 8H), 2.74-3.03 (m, 2H), 3.33-3.83 (m, 2H), 4.45-4.91 (m, 2H), 7.17-8.05 (m, 10H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) rotameric mixture: δ 22.0, 23.3, 23.7, 24.2, 25.0, 36.1, 36.8, 42.4, 42.7, 48.6, 49.2, 54.6, 55.5, 58.1, 125.8, 126.3, 126.6, 128.4, 128.5, 128.7, 129.5, 129.67, 129.74, 129.9, 136.1, 136.4; IR (CHCl<sub>3</sub>) 2942, 1632, 1427 cm<sup>-1</sup>; MS (ESI, MNa<sup>+</sup>, C<sub>22</sub>H<sub>24</sub>N<sub>2</sub>O<sub>2</sub>Na) 371; Anal. Calc'd. for C<sub>22</sub>H<sub>24</sub>N<sub>2</sub>O<sub>2</sub> C 75.83% H 6.94% N 8.04%; Found C 75.66% H 6.94% N 7.93%.

Reduction of the above bisamide (1.15 g, 3.3 mmol) and purification by chromatography (95:5 CH<sub>2</sub>Cl<sub>2</sub>:MeOH) afforded **11e** (0.98 g, 93%) as a waxy white solid: (*R,R*)-**11e** [α]<sub>D</sub><sup>25</sup> = -11.5 (c 1.04, EtOH); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.48-1.75 (m, 8H), 2.44 (d, J=7.7 Hz, 4H), 3.06 (d, J=10.8 Hz, 2H), 3.59 (d, J=13.5 Hz, 2H), 3.72 (d, J=13.5 Hz, 2H), 7.21-7.43 (m, 10H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 16.4, 24.2, 45.6, 56.8, 58.6, 126.7, 128.1, 128.6, 139.8; IR (CHCl<sub>3</sub>) 2964-3022, 2466, 1456 cm<sup>-1</sup>; MS (ESI, MH<sup>+</sup>, C<sub>22</sub>H<sub>29</sub>N<sub>2</sub>) 321.

***N,N'*-bis-Cyclohexylethyl-*cis*-decahydro-1,5-naphthyridine (11f)**. Using compound **10** and cyclohexylacetyl chloride, *N,N'*-cyclohexylacetyl-*cis*-decahydro-1,5-naphthyridine was obtained. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 0.88-0.99 (m), 1.09-1.18 (m), 1.23-1.31 (m), 1.43-1.84 (m), 2.00-2.27 (m), 2.50-2.59 (m), 2.96-3.06 (m), 3.65-3.87 (m), 4.53-4.80 (m); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 21.8, 22.1, 23-27 (m), 33-34 (m), 35-37 (m), 40-42 (m), 48.1, 48.8, 53.0, 54.5, 171-172 (m); MS (ESI, MNa<sup>+</sup>, C<sub>24</sub>H<sub>40</sub>N<sub>2</sub>O<sub>2</sub>Na) 411.6.

Reduction of the above bisamide afforded **11f** (80% for two steps) after chromatography: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.26-1.40 (m, 2H), 1.51-1.68 (m, 8H), 2.56-2.64 (t, J = 9.0 Hz, 2H) 2.68 (s, 2H) 3.04-3.11 (d, J = 9.0 Hz, 2H) MS (ESI, MH<sup>+</sup>, C<sub>24</sub>H<sub>45</sub>N<sub>2</sub>) 361.6.

***N,N'*-bis-Trityl-*cis*-decahydro-1,5-naphthyridine (11g).** Diamine **10** (50 mg, 0.36 mmol), trityl bromide (254 mg, 0.78 mmol), and Et<sub>3</sub>N (109  $\mu$ l, 0.78 mmol) were mixed in CH<sub>2</sub>Cl<sub>2</sub> (52.0 mL) at rt for 1 d. The reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with sat'd aqueous NaHCO<sub>3</sub>, and dried over Na<sub>2</sub>SO<sub>4</sub>. After concentrating in vacuo, the crude material was suspended in Et<sub>2</sub>O, filtered and washed with more Et<sub>2</sub>O. The resultant solid (47%) was pure **11g**: <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  0.35 (d, J = 14.5 Hz, 2H), 1.15 (m, 4H), 1.35 (m, 2H), 2.36 (dt, J = 3.0, 12.3 Hz, 2H), 2.74 (d, J = 12.0 Hz, 2H), 3.69 (d, J = 12.2 Hz, 2H), 7.21 (m, 22H), 7.47 (d, J = 6.9 Hz, 8H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  18.7, 25.2, 40.9, 55.4, 125.8, 127.5, 127.9, 129.6, 145.4.

**General Procedure for Lithiation-Substitution of *N*-Boc-Pyrrolidine (Figure 6).** To the diamine ligand (1.3 equiv) in Et<sub>2</sub>O (0.3-0.1 M) at -78 °C was added sBuLi (1.3 equiv). The reaction mixture was stirred at -78 °C for 10 min and **12** (1.0 equiv) was added. The mixture was kept at -78 °C for 6 h. An aliquot was taken out and quenched by MeOD to assay for anion formation. The D-exchange ratio was determined by <sup>1</sup>H NMR. The remainder of the reaction mixture was then quenched by TMSCl. The resultant mixture was warmed up to rt slowly. Workup consisted of addition of water, extraction of the aqueous layer with Et<sub>2</sub>O, washing the combined Et<sub>2</sub>O layers with 1N HCl, drying over MgSO<sub>4</sub>, filtration and concentration *in vacuo* to obtain crude **13** which can be further purified by flash chromatography (1/25 = EtOAc/hexane) to give pure **13** as a colorless oil. Analytical data was identical to that reported by Beak (see references in main text). The enantiomeric purity and conversion were determined using a Cylcodex B column (95 °C isothermal, 10 psi)  $t_R$  (S) = 86.3 min,  $t_R$  (R) = 87.6 min.

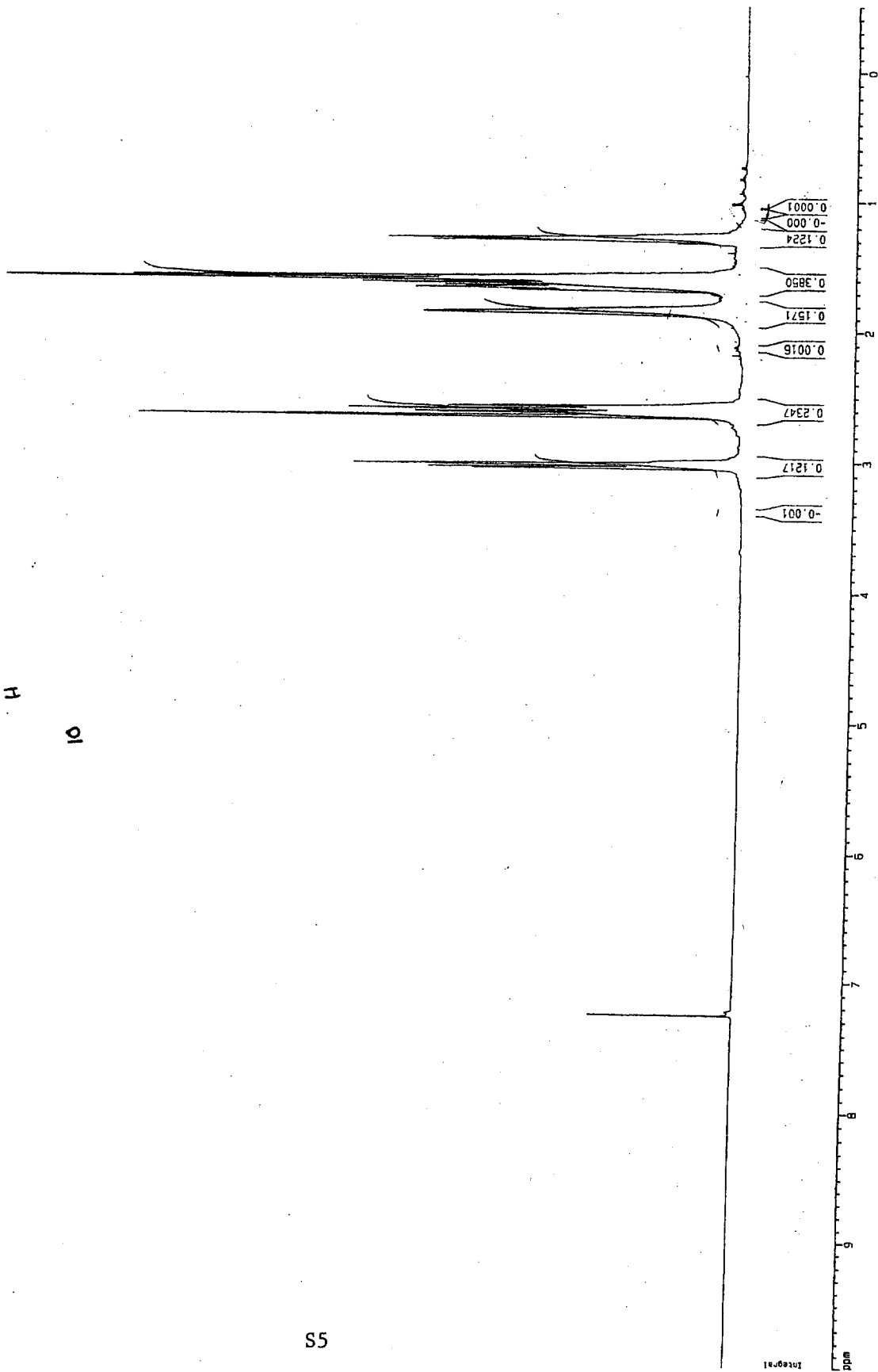
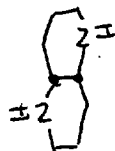
**General Procedure for Lithiation-Substitution of *N,N'*-Diisopropyl-*o*-ethylbenzamide (Figure 7).** To a solution of ligand (0.55 mmol) in *n*-pentane/MTBE = 1/1 (15 mL) or *n*-pentane at -78 °C was added sBuLi (0.55 mmol). The mixture was stirred for 10 min and then transferred to a precooled solution of **14** in the same solvent (15 mL). The resulting purple mixture was kept at -78 °C for 1.5 h (the warm-up protocol would involve 30 min at -78 °C, 1 h at -20 °C and recooling to -78 °C prior to adding the electrophile). The electrophile (1.5 equiv) was added (distilled acetone for **15a** and allyl-X for **15b**). After the solution became colorless, MeOH was added. The solution was then diluted with 1N HCl and extracted with Et<sub>2</sub>O. The combined Et<sub>2</sub>O extracts were dried over MgSO<sub>4</sub>, filtered, and concentrated *in vacuo*. The resultant mixture which was chromatographed with 20% EtOAc/hexane to yield pure **15a** and with 8% EtOAc/hexane to yield pure **15b**. The analytical data for **7** and **8** proved identical to that reported by Beak (see references in main text). For **15a**, enantiomeric purity was determined by HPLC using a Chiralpak AD column with iPrOH/hexanes/Et<sub>2</sub>NH (7.5:92.5:0.1) at 1.0 mL/min ( $t_R$  (R) = 8.1 min,  $t_R$  (S) = 9.5 min). For **15b**, enantiomeric purity was determined by HPLC using a semi-prep Chiralpak AD column with iPrOH/hexanes (2.5:97.5) at 2.4 mL/min ( $t_R$  (S) = 15.0 min,  $t_R$  (R) = 17.6 min).

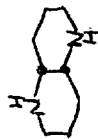
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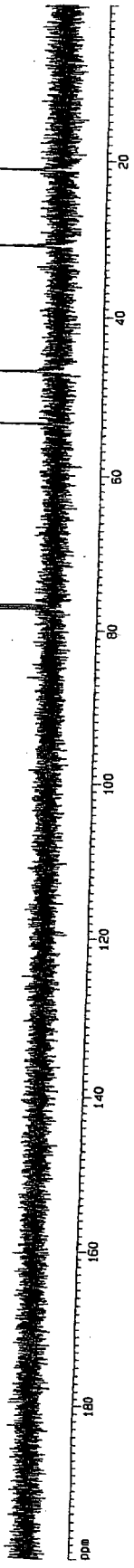
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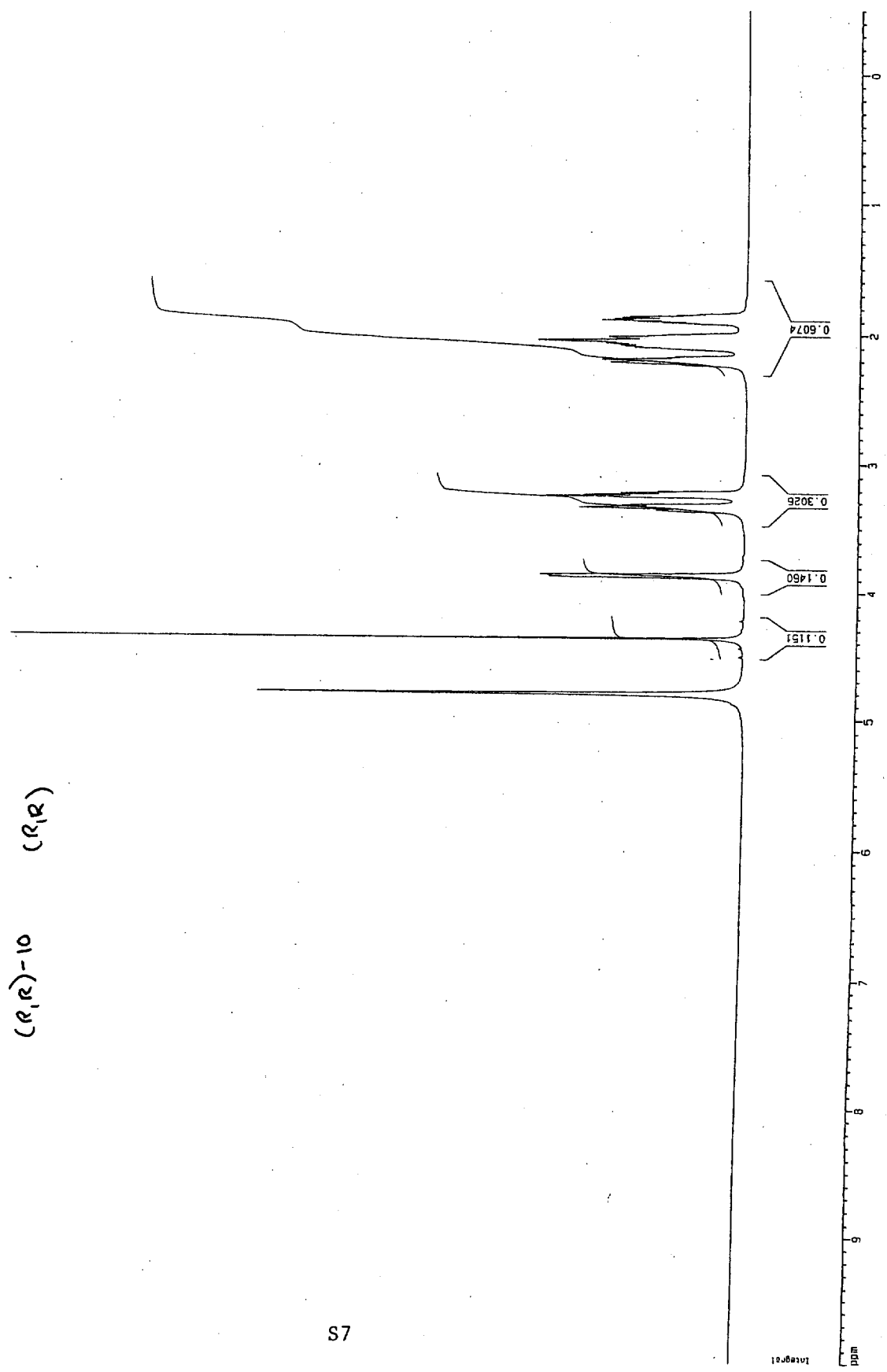
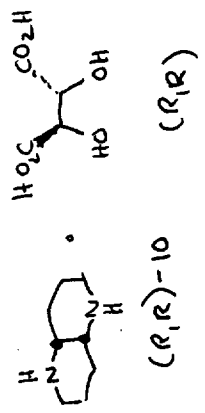
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EXPNO 1  
PROCNO 1

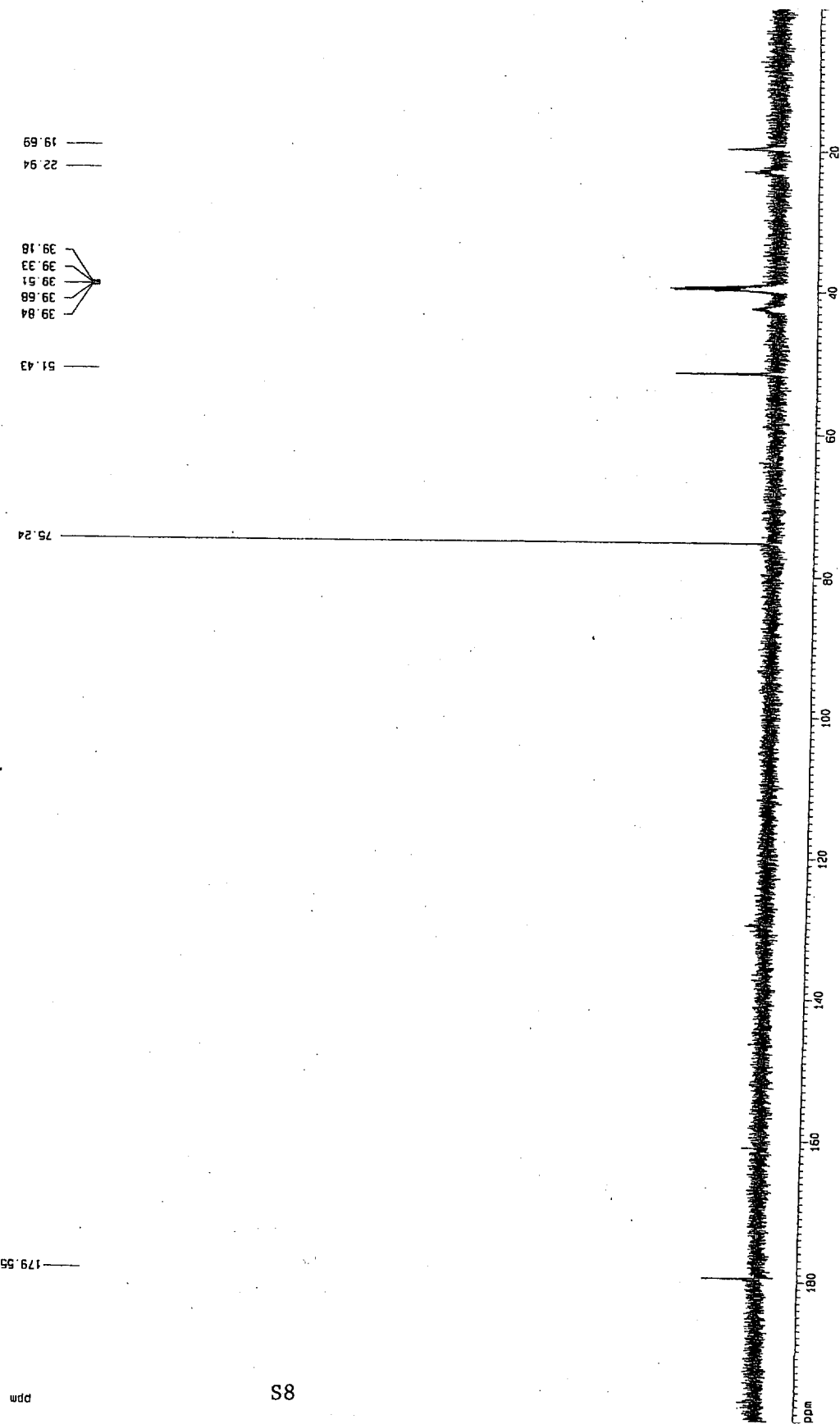
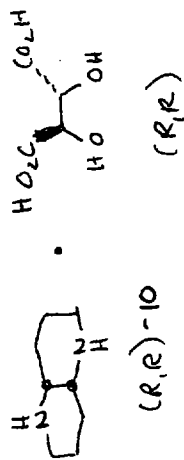
F2 - Acquisition Parameters  
Date\_ 980918  
Time 10.58  
INSTRUM spect  
PROBHD 5 mm QNP 1H  
PULPROG zgpg30  
SOLVENT DMSO-d6  
NS 128  
DS 2  
OS 2  
SH 3333.332 Hz  
FIDRES 0.508265 Hz  
AQ 0.9826500 sec  
RG 16384  
DE 15.000 umsc  
TE 300.2 umsc  
TD 32768  
SFO1 400.146 MHz  
NUC1 13C  
SFO2 101.253 MHz  
NUC2 13C  
SFO3 125.762148 MHz  
SFO4 125.762148 MHz  
SFO5 125.762148 MHz  
SFO6 125.762148 MHz  
SFO7 125.762148 MHz  
SFO8 125.762148 MHz  
SFO9 125.762148 MHz  
SFO10 125.762148 MHz  
SFO11 125.762148 MHz  
SFO12 125.762148 MHz  
SFO13 125.762148 MHz  
SFO14 125.762148 MHz  
SFO15 125.762148 MHz  
SFO16 125.762148 MHz  
SFO17 125.762148 MHz  
SFO18 125.762148 MHz  
SFO19 125.762148 MHz  
SFO20 125.762148 MHz  
SFO21 125.762148 MHz  
SFO22 125.762148 MHz  
SFO23 125.762148 MHz  
SFO24 125.762148 MHz  
SFO25 125.762148 MHz  
SFO26 125.762148 MHz  
SFO27 125.762148 MHz  
SFO28 125.762148 MHz  
SFO29 125.762148 MHz  
SFO30 125.762148 MHz  
SFO31 125.762148 MHz  
SFO32 125.762148 MHz  
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SFO34 125.762148 MHz  
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SFO37 125.762148 MHz  
SFO38 125.762148 MHz  
SFO39 125.762148 MHz  
SFO40 125.762148 MHz  
SFO41 125.762148 MHz  
SFO42 125.762148 MHz  
SFO43 125.762148 MHz  
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SFO45 125.762148 MHz  
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SFO70 125.762148 MHz  
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SFO76 125.762148 MHz  
SFO77 125.762148 MHz  
SFO78 125.762148 MHz  
SFO79 125.762148 MHz  
SFO80 125.762148 MHz  
SFO81 125.762148 MHz  
SFO82 125.762148 MHz  
SFO83 125.762148 MHz  
SFO84 125.762148 MHz  
SFO85 125.762148 MHz  
SFO86 125.762148 MHz  
SFO87 125.762148 MHz  
SFO88 125.762148 MHz  
SFO89 125.762148 MHz  
SFO90 125.762148 MHz  
SFO91 125.762148 MHz  
SFO92 125.762148 MHz  
SFO93 125.762148 MHz  
SFO94 125.762148 MHz  
SFO95 125.762148 MHz  
SFO96 125.762148 MHz  
SFO97 125.762148 MHz  
SFO98 125.762148 MHz  
SFO99 125.762148 MHz  
SFO100 125.762148 MHz



ppm

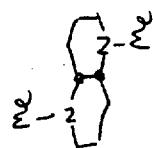


Current Data Parameters  
 NAME II-1281rvrat  
 EXPNO 323  
 PROCNO 1  
 F2 - Acquisition Parameters  
 Date\_ 090919  
 Time 13.32  
 INSTRUM emx500  
 PROBM0 5 nu Dual 13  
 PULPROG zgpg30  
 TD 32768  
 SOLVENT DMS-D6  
 NS 47  
 DS 4  
 SWH 6724.056 Hz  
 FIDRES 0.163814 Hz  
 AQ 2.119750494 sec  
 RG 256  
 ON 83.000 MHz  
 DE 118.57 MHz  
 TE 300.0 K  
 FL1 1.0000000 sec  
 FL2 1.0000000 sec  
 DE 118.57 MHz  
 SF01 500.362510 MHz  
 NUCLEUS 1H  
 F2 - Processing parameters  
 SI 163814  
 SF 500.362510 MHz  
 SW 6724.056 MHz  
 SN 3203.80 Hz  
 LB 0.500 Hz  
 GB -250.15 Hz  
 PC 0.30000 cm  
 WDM 150.115590 Hz/c  
 ID 1000 pilot parameters  
 CR 35.00 cm  
 CP 10.00 cm  
 F1 3003.80 Hz  
 F2 0.500 cm  
 F3 -250.15 Hz  
 PPM0 0.30000 cm  
 WDM 150.115590 Hz/c

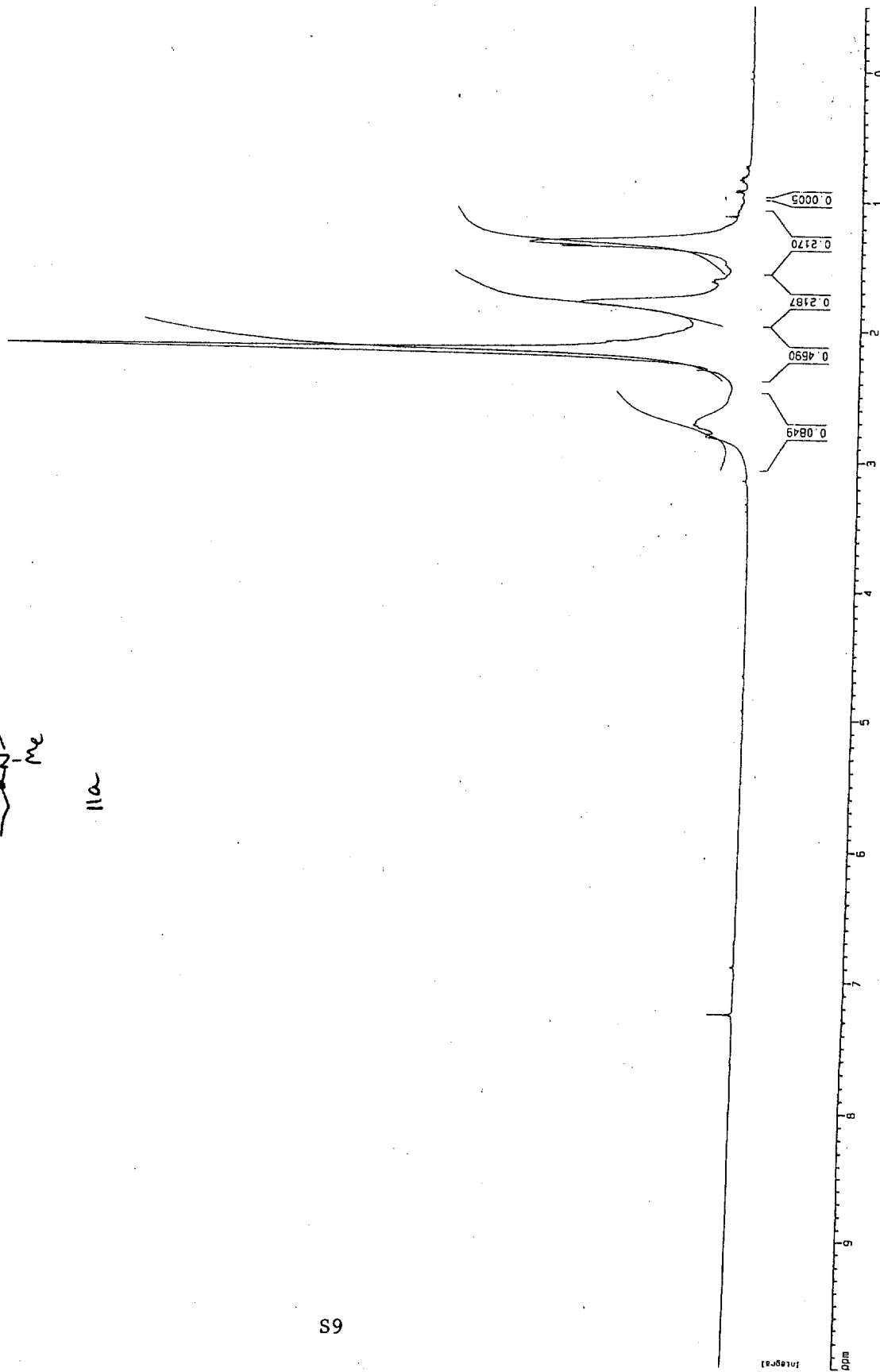


Current Data Parameters  
 Name 11-12Carbon  
 ELEM C  
 PROCN 1  
  
 F2 - Acquisition Parameters  
 Date\_ 990919  
 Time 15.09  
 INSTRUM 5  
 PDBNO 5  
 TO PRG0 099.917  
 SOLVENT CDCl3  
 NS 754  
 DS 2  
 SFO 3333.132 Hz  
 FIDRES 0.308256 Hz  
 AQ 0.392990 sec  
 RG 6334  
 DE 18.75 usec  
 TE 300.0 K  
 H1 40 dB  
 S4 24 dB  
 O1 0.0200000 sec  
 D1 2.0000000 sec  
 S1 107.10 usec  
 S2 18 dB  
 P1 3.00 usec  
 DE 18.75 usec  
 SFO1 125.938148 MHz  
 NUC16 13C  
  
 F2 - Processing Parameters  
 SI 20758  
 SF 125.932948 MHz  
 NU 0  
 NDN 0  
 LB 0.00 Hz  
 GB 0  
 PC 1.40  
  
 1D NMR 0101 parameters  
 CX 35.00 Co  
 FIP 200.000 Dpa  
 F1 25164.50 Hz  
 F2 0.000 Dpa  
 HZ 140.000 Hz  
 PRNOM 5 71429 000A1  
 NUCM 719.99553 Hz/G1





11a



Current Data Parameters  
 NAME 11-22801.mv  
 EXPNO 1  
 PROCNO 1  
 F2 - Acquisition Parameters  
 Date\_ 20000303  
 Time 12:00  
 INSTRUM spect  
 PROBHD 5 mm QNP 1H  
 PULPROG zgpg30  
 TD 32768  
 SOLVENT CDCl3  
 NS 256  
 DS 4  
 SWH 6024.096 Hz  
 FIDRES 0.148417 Hz  
 AQ 2.7197540 sec  
 RG 328  
 DM 83.000 usec  
 DE 118.57 usec  
 TE 300.0 K  
 HL 1.00  
 H1 1.0000000 sec  
 D1 12.00 usec  
 DE 118.57 usec  
 SFO1 500.362518 MHz  
 NUC1 1H  
 F2 - Processing parameters  
 SI 32768  
 SF 500.3600176 MHz  
 RG 328  
 SSB no  
 LB 0.00 Hz  
 GB 0  
 PC 1.40  
 F0 0.00000000  
 CK 35.00 cm  
 F1 10.000 cm  
 F2 10.000 cm  
 F3 -4.500 cm  
 F4 -250.20 Hz  
 PPM0 0.30000 ppm  
 HZ0 150.11700 Hz/c

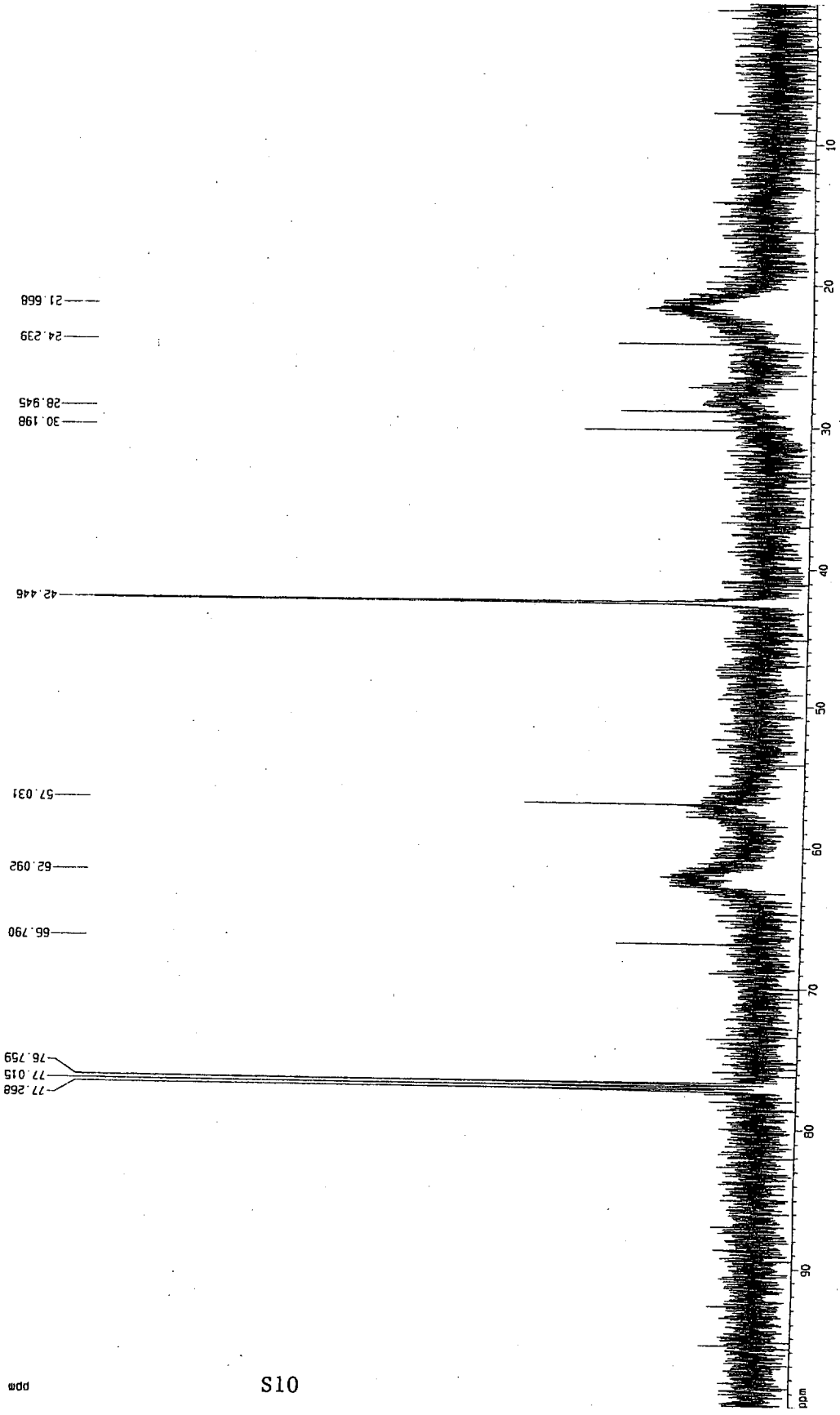
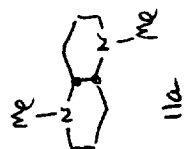
```

Current Data Parameters
NAME      11-226a.mn
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    991200
Time     14:16
INSTRUM  spect
PROBHD   5 mm QNP 1H
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       2
DS       2
SWH      3333.32 Hz
FIDRES   0.50622 Hz
AQ       0.393200 sec
RG       16384
DM       15.000 usec
DE       19.75 usec
TE       300.0 K
AQ1      40.00 usec
AQ2      40.00 usec
AQ3      40.00 usec
SFO1     125.830448 MHz
SFO2     125.830448 MHz
MAGLEV   13C

F2 - Processing parameters
SI       32768
SF       125.831767 MHz
RG       1024
SSB      0
LB       0.00 Hz
GB       0
PC       1.40

1D NMR plot parameters
SI       32768
SF       125.831767 MHz
RG       1024
SSB      0
LB       0.000000 MHz
F2P      0.000000 MHz
F2       0.00 Hz
PPMCH    2.85714 ppm/Hz
MCHN     359.49478 Hz/Hz
    
```



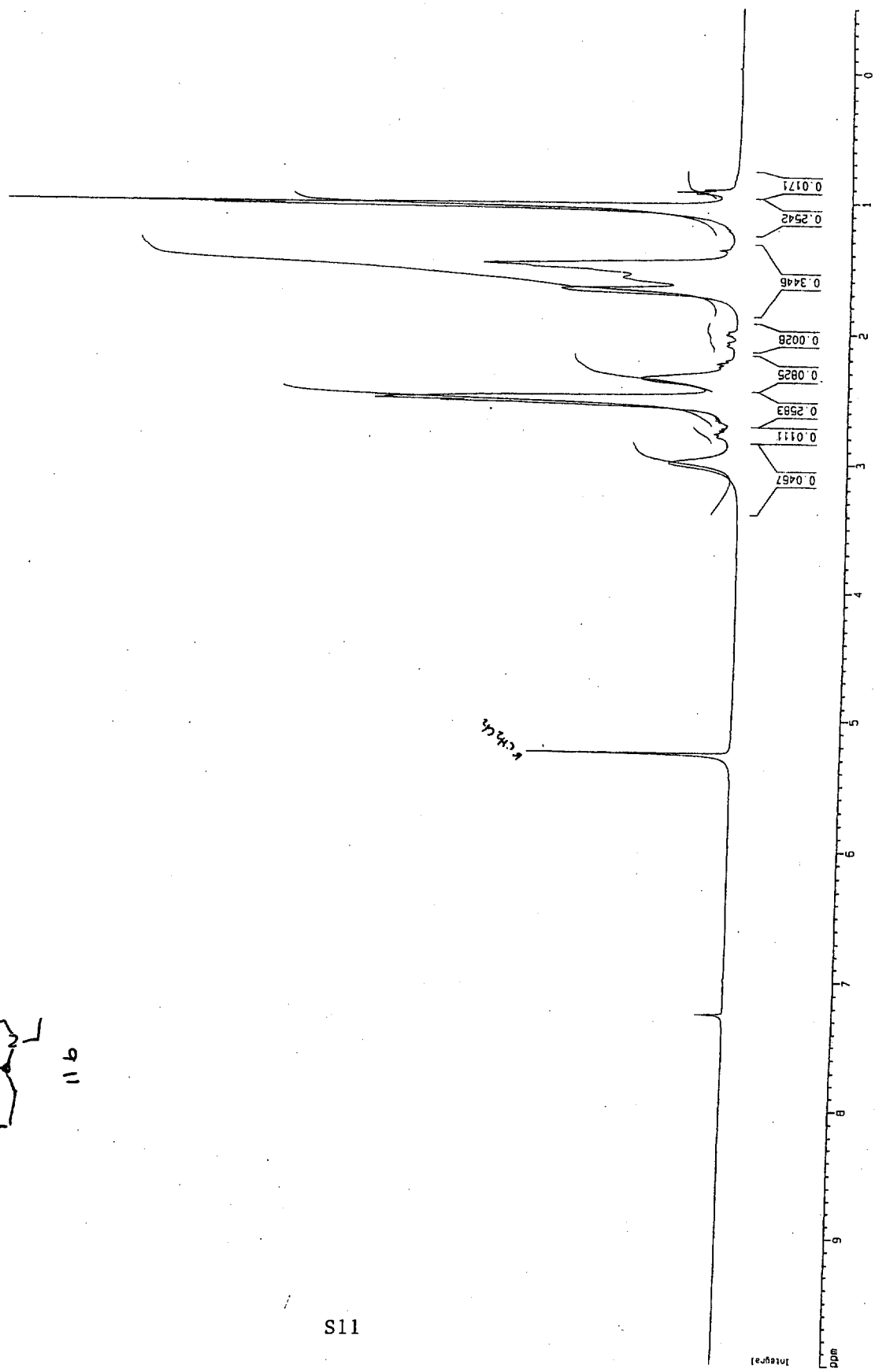


Current Data Parameters  
 NAME III-23  
 EXPNO 1  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 9/09/00  
 Time 16:13  
 INSTRUM amc500  
 PROBHD 5 mm Dual 13  
 PULPROG zg  
 TD 32768  
 SOLVENT CDCl3  
 CS 28  
 SFO1 500.3522518 MHz  
 NUC1 13  
 SFO2 101.628125 MHz  
 NUC2 13  
 SFO3 500.3522518 MHz  
 NUC3 1H

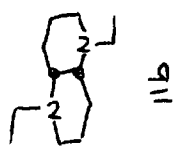
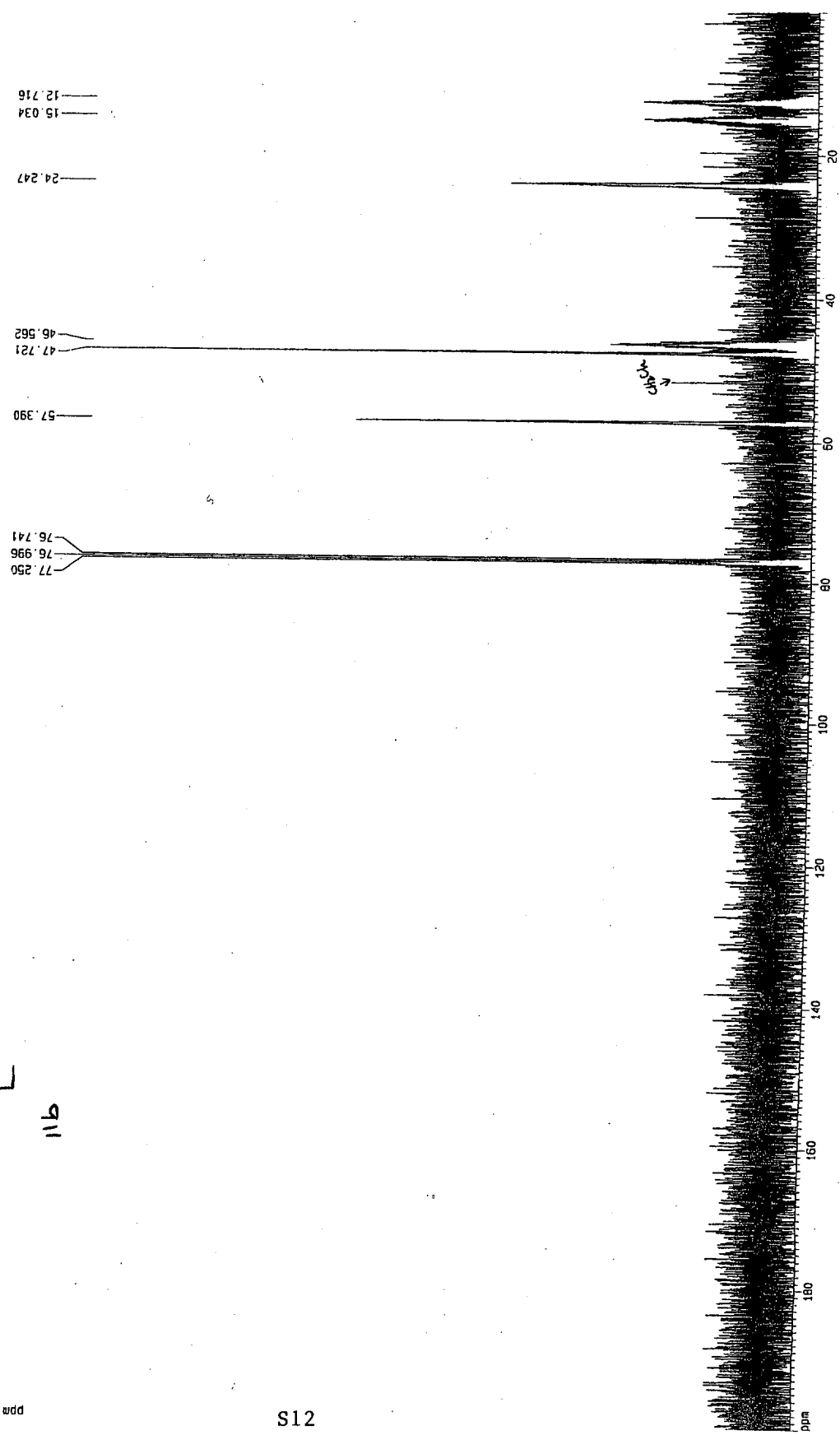
F2 - Processing parameters  
 SI 16384  
 SF 500.3500161 MHz  
 WDW m  
 SSB 0  
 LB 0.00 Hz  
 GB 0  
 PC 1.40

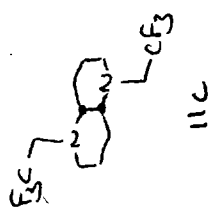
ID MR plot parameters  
 FI 35.00 ca  
 F1 5003.90 Hz  
 F2 -0.500 ppm  
 FWHM 0.30000 ppm  
 HZ 150.11700 Hz/C



```

Current Data Parameters
Name: 111-23a
Date_: 1
PROCNO: 1
F2 - Acquisition Parameters
Date_: 5/06/00
Time: 16.25
INSTRUM: AMX500
PROBHD: 5 mm QNP 1H
PULPROG: zgpg30
PCPDPR: 1
SOLVENT: CDCl3
NS: 2
DS: 4
AQ: 3.3333333 Hz
FIDRES: 0.5000000 Hz
AQRES: 0.9630000 sec
RG: 327.68
RGRES: 1.9000000 sec
TE: 300.0 K
NUC1: 13C
NUC2: 13C
NUC3: 13C
NUC4: 13C
NUC5: 13C
NUC6: 13C
NUC7: 13C
NUC8: 13C
NUC9: 13C
NUC10: 13C
NUC11: 13C
NUC12: 13C
NUC13: 13C
NUC14: 13C
NUC15: 13C
NUC16: 13C
NUC17: 13C
NUC18: 13C
NUC19: 13C
NUC20: 13C
NUC21: 13C
NUC22: 13C
NUC23: 13C
NUC24: 13C
NUC25: 13C
NUC26: 13C
NUC27: 13C
NUC28: 13C
NUC29: 13C
NUC30: 13C
NUC31: 13C
NUC32: 13C
NUC33: 13C
NUC34: 13C
NUC35: 13C
NUC36: 13C
NUC37: 13C
NUC38: 13C
NUC39: 13C
NUC40: 13C
NUC41: 13C
NUC42: 13C
NUC43: 13C
NUC44: 13C
NUC45: 13C
NUC46: 13C
NUC47: 13C
NUC48: 13C
NUC49: 13C
NUC50: 13C
F2 - Processing Parameters
SI: 125.0231756 MHz
SF: 125.0231756 MHz
WDW: EM
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 1.40
SFO: 125.0231756 MHz
AQ: 3.3333333 Hz
RG: 327.68
RGRES: 1.9000000 sec
TE: 300.0 K
NUC1: 13C
NUC2: 13C
NUC3: 13C
NUC4: 13C
NUC5: 13C
NUC6: 13C
NUC7: 13C
NUC8: 13C
NUC9: 13C
NUC10: 13C
NUC11: 13C
NUC12: 13C
NUC13: 13C
NUC14: 13C
NUC15: 13C
NUC16: 13C
NUC17: 13C
NUC18: 13C
NUC19: 13C
NUC20: 13C
NUC21: 13C
NUC22: 13C
NUC23: 13C
NUC24: 13C
NUC25: 13C
NUC26: 13C
NUC27: 13C
NUC28: 13C
NUC29: 13C
NUC30: 13C
NUC31: 13C
NUC32: 13C
NUC33: 13C
NUC34: 13C
NUC35: 13C
NUC36: 13C
NUC37: 13C
NUC38: 13C
NUC39: 13C
NUC40: 13C
NUC41: 13C
NUC42: 13C
NUC43: 13C
NUC44: 13C
NUC45: 13C
NUC46: 13C
NUC47: 13C
NUC48: 13C
NUC49: 13C
NUC50: 13C
IO MSG plot parameters
CX: 30.00 cm
FIP: 200.000 ppm
F1: 25164.63 Hz
F2: 0.000 ppm
F3: 0.000 Hz
PROCH: 13C
MORPH: 718.9858 Hz
    
```



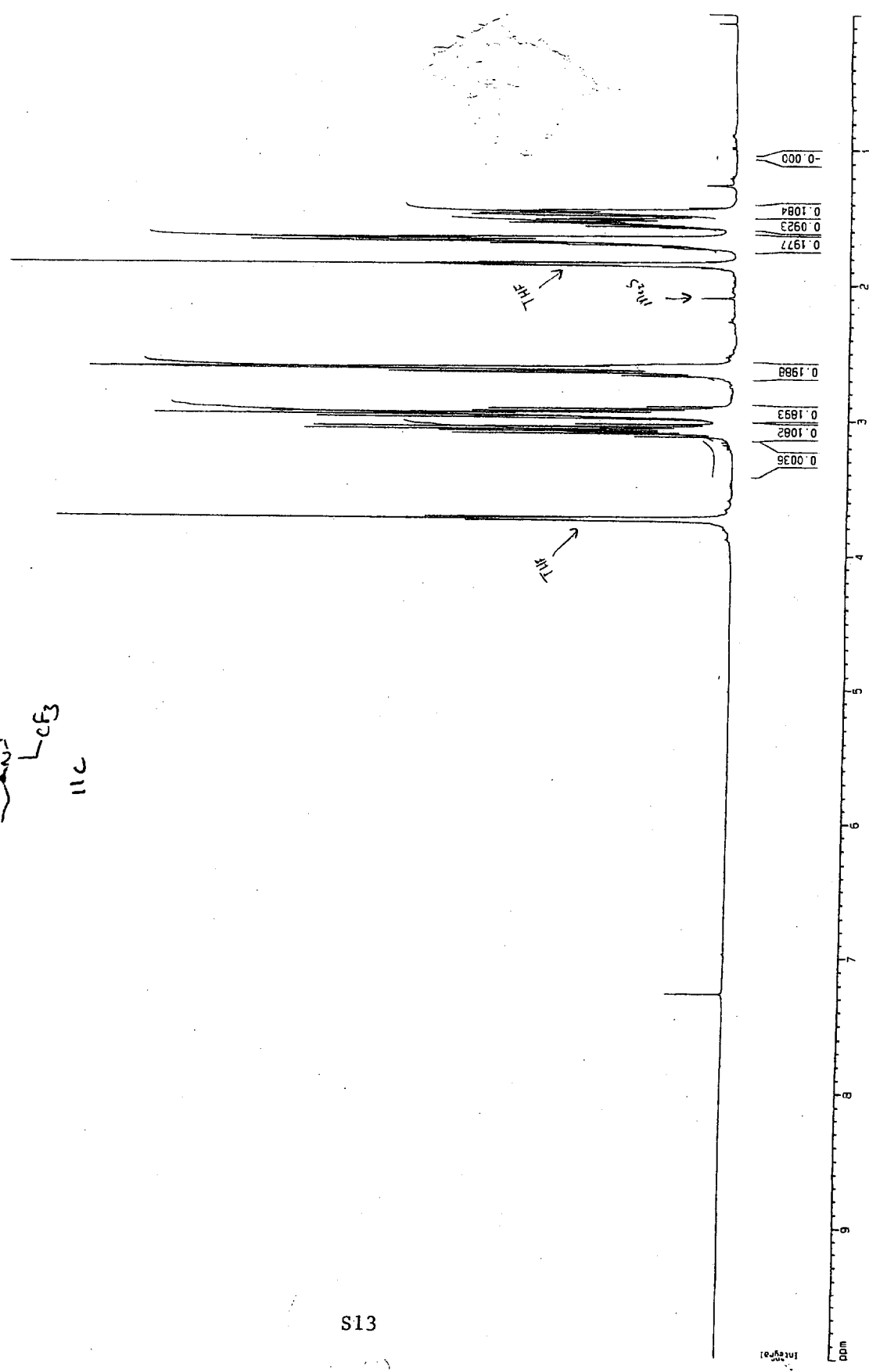


Current Data Parameters  
 NAME 11-177  
 EXPNO 1  
 PROCNO 1

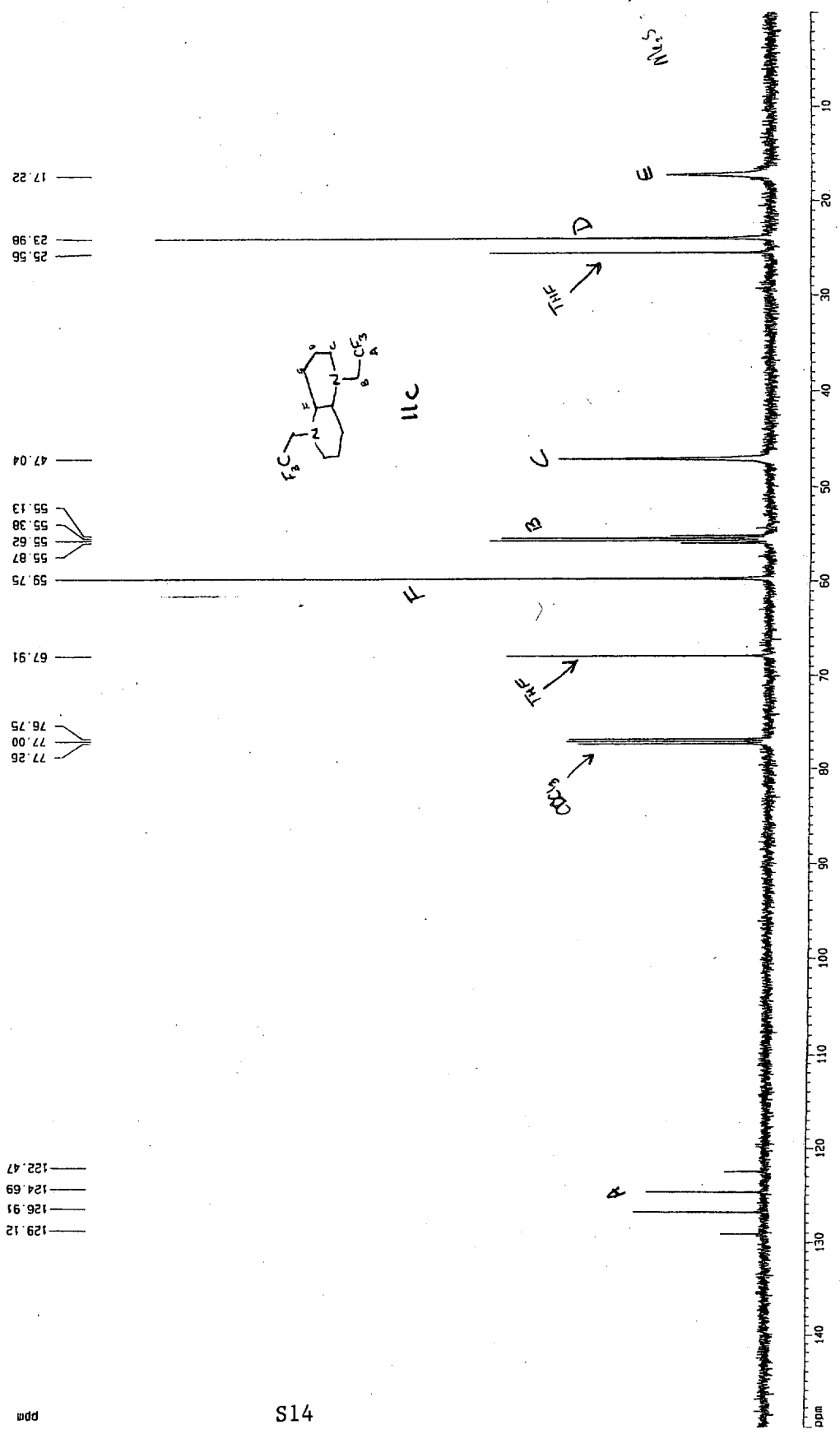
F2 - Acquisition Parameters  
 FILE\_ 991019  
 F2 300  
 INSTRUM spect  
 PULPROG zgpg30  
 S wa Dn1 13  
 TO 32768  
 SOLVENT CDCl3  
 NS 40  
 DS 4  
 SH 0624.056 Hz  
 FWHM 0.163841 Hz  
 AQ 2.7151740 sec  
 RG 65  
 DM 83.000 usec  
 DE 118.97 usec  
 TE 300.0 K  
 HL1 1.0000000 sec  
 O1 12.00 usec  
 DE 12.00 usec  
 O2 12.00 usec  
 DE 12.00 usec  
 SF01 500.3622710 MHz  
 NUCLEUS 1H

F2 - Processing parameters  
 SI 16394  
 SF 500.3600058 MHz  
 EN  
 GB 0.10 Hz  
 LB 0.0  
 PC 1.40

1D NMR plot parameters  
 CX 35.00 cps  
 FIP 10.000 cps  
 F2P 500.350 Hz  
 F2 300.00 MHz  
 F2 0.00 Hz  
 FREQM 0.285710000 MHz  
 NUCM 142.95808 Hz/c



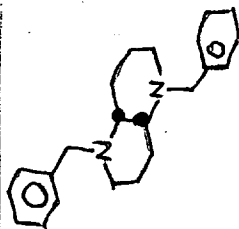
Current Data Parameters  
 NAME 11-177carbon  
 EXPNO 1  
 PROCNO 1  
 F2 - Acquisition Parameters  
 INSTRUM spect  
 PROBRG zgpg30  
 PULPROG zgpg30  
 SOLVENT CDCl3  
 NS 74  
 DS 4  
 SWH 3333.332 Hz  
 FIDRES 0.56826 Hz  
 AQ 0.0300000 sec  
 RG 6384  
 DM 15.000 usec  
 DE 18.75 usec  
 TE 300.0 K  
 F1 101.626130 MHz  
 F2 101.626130 MHz  
 SFO1 125.832148 MHz  
 NUC1 13C  
 F2 - Processing parameters  
 SI 32768  
 SF 125.832148 MHz  
 SN 0  
 SS 0  
 SSB 0  
 GB 0  
 PC 1.40  
 RB 0  
 LB 1.50 Hz  
 GB 0  
 TO 15.000 sec  
 DS 4  
 SWH 3333.332 Hz  
 FIDRES 0.56826 Hz  
 AQ 0.0300000 sec  
 RG 6384  
 DM 15.000 usec  
 DE 18.75 usec  
 TE 300.0 K  
 F1 101.626130 MHz  
 F2 101.626130 MHz  
 SFO1 125.832148 MHz  
 NUC1 13C  
 F2 - Acquisition Parameters  
 INSTRUM spect  
 PROBRG zgpg30  
 PULPROG zgpg30  
 SOLVENT CDCl3  
 NS 74  
 DS 4  
 SWH 3333.332 Hz  
 FIDRES 0.56826 Hz  
 AQ 0.0300000 sec  
 RG 6384  
 DM 15.000 usec  
 DE 18.75 usec  
 TE 300.0 K  
 F1 101.626130 MHz  
 F2 101.626130 MHz  
 SFO1 125.832148 MHz  
 NUC1 13C  
 F2 - Processing parameters  
 SI 32768  
 SF 125.832148 MHz  
 SN 0  
 SS 0  
 SSB 0  
 GB 0  
 PC 1.40  
 RB 0  
 LB 1.50 Hz  
 GB 0  
 TO 15.000 sec  
 DS 4  
 SWH 3333.332 Hz  
 FIDRES 0.56826 Hz  
 AQ 0.0300000 sec  
 RG 6384  
 DM 15.000 usec  
 DE 18.75 usec  
 TE 300.0 K  
 F1 101.626130 MHz  
 F2 101.626130 MHz  
 SFO1 125.832148 MHz  
 NUC1 13C



11c

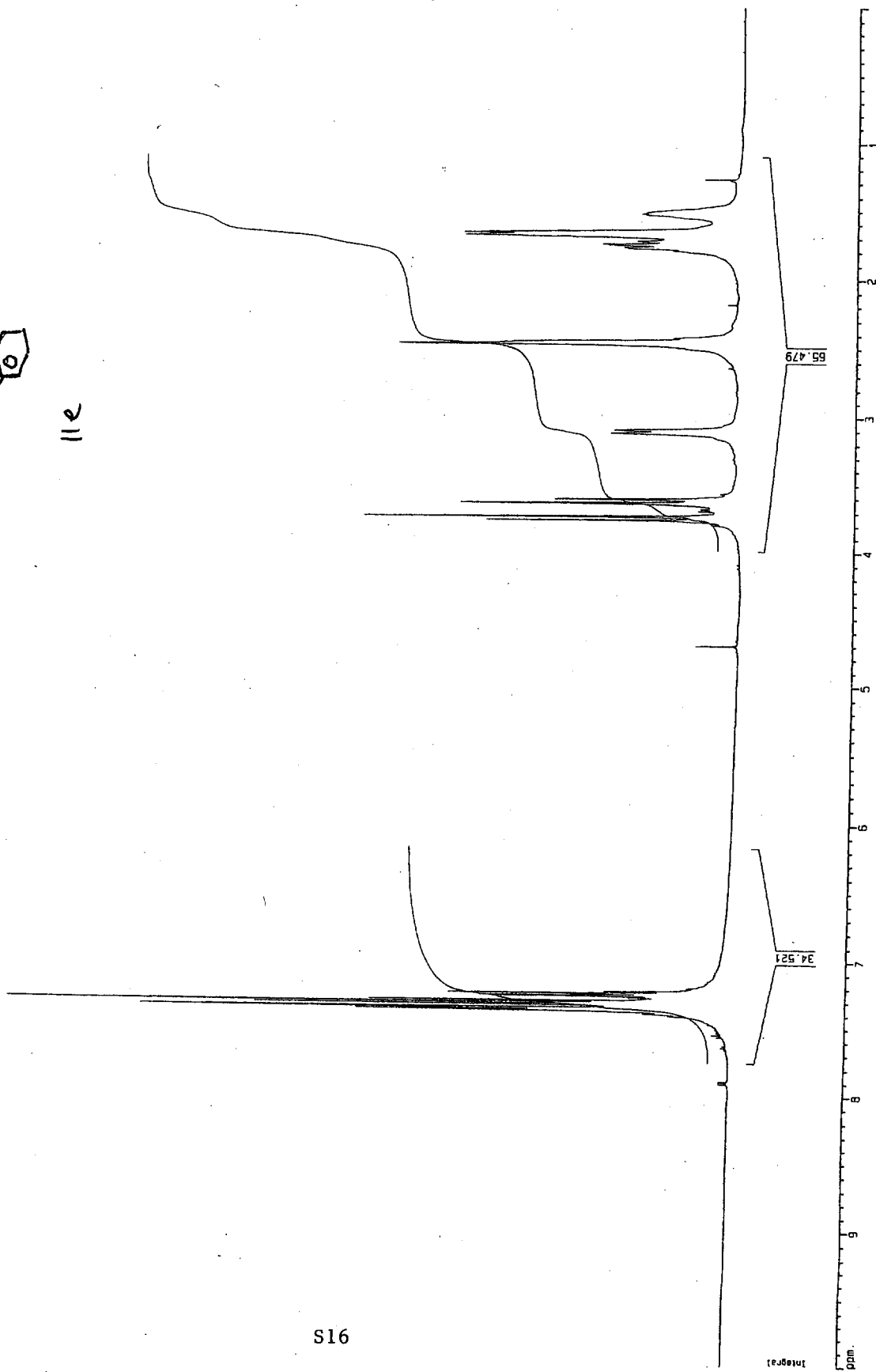
ppm





11e

Current Data Parameters  
NAME: benzylamine  
EXPNO: 1  
PROCNO: 1  
F2 - Acquisition Parameters  
Date\_ : 9/07/95  
Time : 18.21  
INSTRUM : am500  
PROBHD : 5 mm Dual 13  
PULPROG : zgpg30  
TD : 32768  
SOLVENT : CDCl3  
AQ : 3.23  
RG : 320  
SM : 6024.066 Hz  
FIDRES : 0.188841 Hz  
AQ : 2.7197540 sec  
RG : 128  
DM : 93.000 usec  
DE : 118.57 usec  
TE : 300.0 K  
D1 : 1.0000000 sec  
D11 : 0.0000000 sec  
D2 : 12.00 usec  
DE : 118.57 usec  
SFO1 : 500.362518 MHz  
NAME : MAALEUS  
F2 - Processing parameters  
SI : 10284  
SF : 500.362518 MHz  
WDW : em  
SSB : 0  
LB : 0.00 Hz  
GB : 0  
PC : 1.40  
AQ : 3.23000000  
RG : 128  
FIR : 10.000 usec  
F1 : 5003.300 Hz  
F2 : 0.000 usec  
F3 : 0.00 Hz  
PPhOH : 0.8871 ppm/  
HZOH : 142.8655 Hz/c







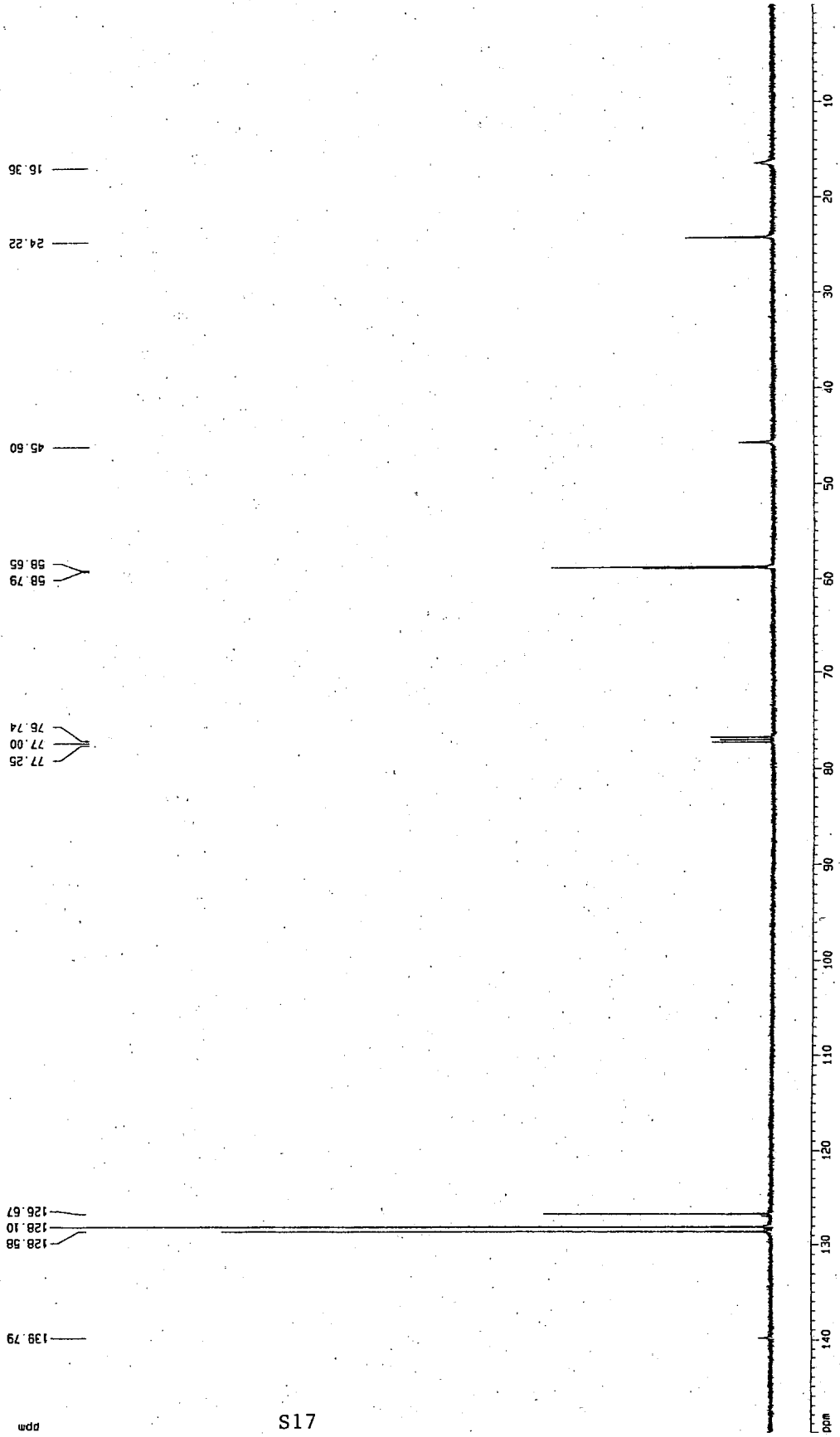
11e

Current Data Parameters  
 NAME: Denty(1)labline  
 EXPNO: 1  
 PROCNO: 1

F2 - Acquisition Parameters  
 Date\_: 5/9/29  
 Time: 16.36  
 INSTRUM: spect  
 PROBR00: 9.40 Data 13  
 PULPROG: zgpg, q14  
 TD: 65536  
 SOA, VENT: CXC13  
 NS: 133  
 DS: 2  
 SWH: 3333.332 Hz  
 FIDRES: 0.504256 Hz  
 AQ: 0.983000 SEC  
 RG: 16384  
 DM: 15.000 usec  
 DE: 19.75 usec  
 TE: 300.0 K  
 N1: 40.00  
 S1: 24.81  
 0.03000000 Hz  
 CPDPRG: wall216  
 P31: 107.00 usec  
 P1: 5.00 usec  
 P2: 18.00 usec  
 SFO1: 125.632148 MHz  
 NUC16S: 13C

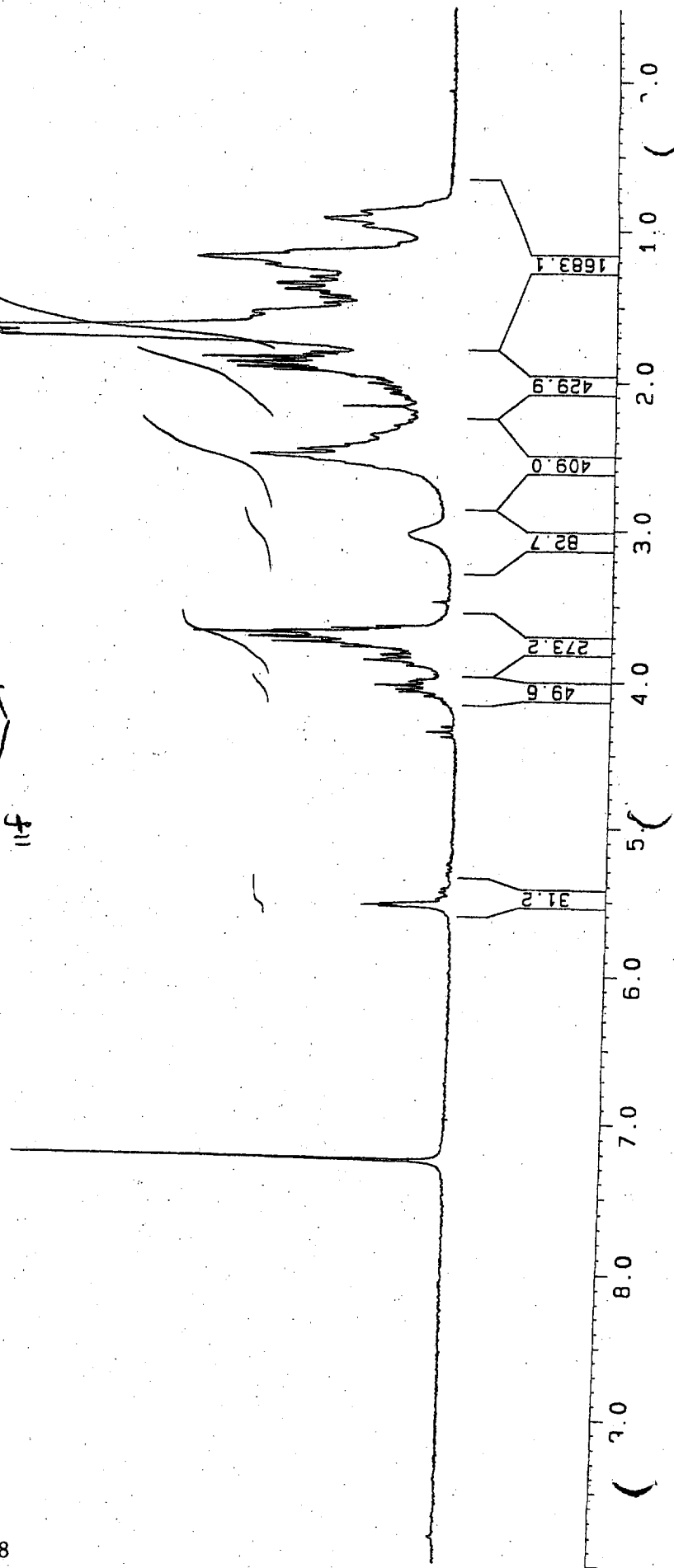
F2 - Processing parameters  
 SI: 32768  
 SF: 125.621777 MHz  
 W: 11  
 SSB: 0  
 LB: 0.00 Hz  
 DB: 0  
 PC: 1.40

ID list parameters  
 C1: 13.00 usec  
 F1P: 150.000 usec  
 F1: 18873.48 Hz  
 F2P: 0.000 usec  
 F2: 0.00 Hz  
 PPM0: 4.28871 ppm/  
 NZDN: 339.24219 Hz/ci

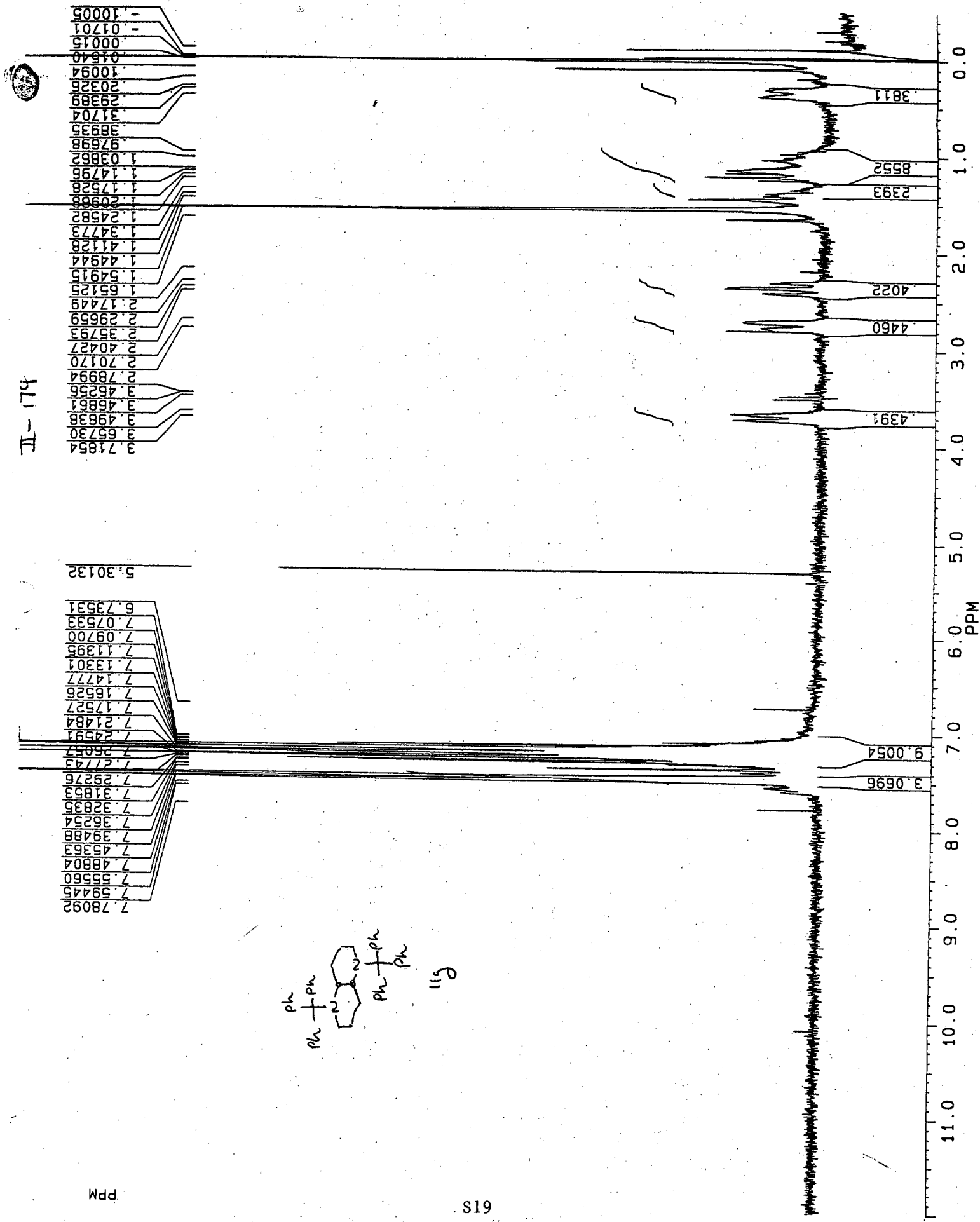


II-67 CDCl3 XL 7/23/99

5.54719
5.53750
5.52743
5.51851
4.05674
4.03104
3.86413
3.74041
3.72983
3.71228
3.70855
3.70037
3.69376
3.68341
3.67372
3.65449
2.52988
2.49719
2.4626
2.42570
2.16844
1.93623
1.92972
1.92240
1.91035
1.90057
1.88973
1.88534
1.88095
1.86513
1.84701
1.82987
1.81497
1.69755
1.68230
1.66563
1.65318
1.54465
1.41607
1.39335
1.35537
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1.23549
1.18009
1.14531
0.91824
0.87243



Mdd

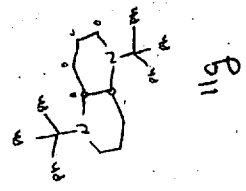
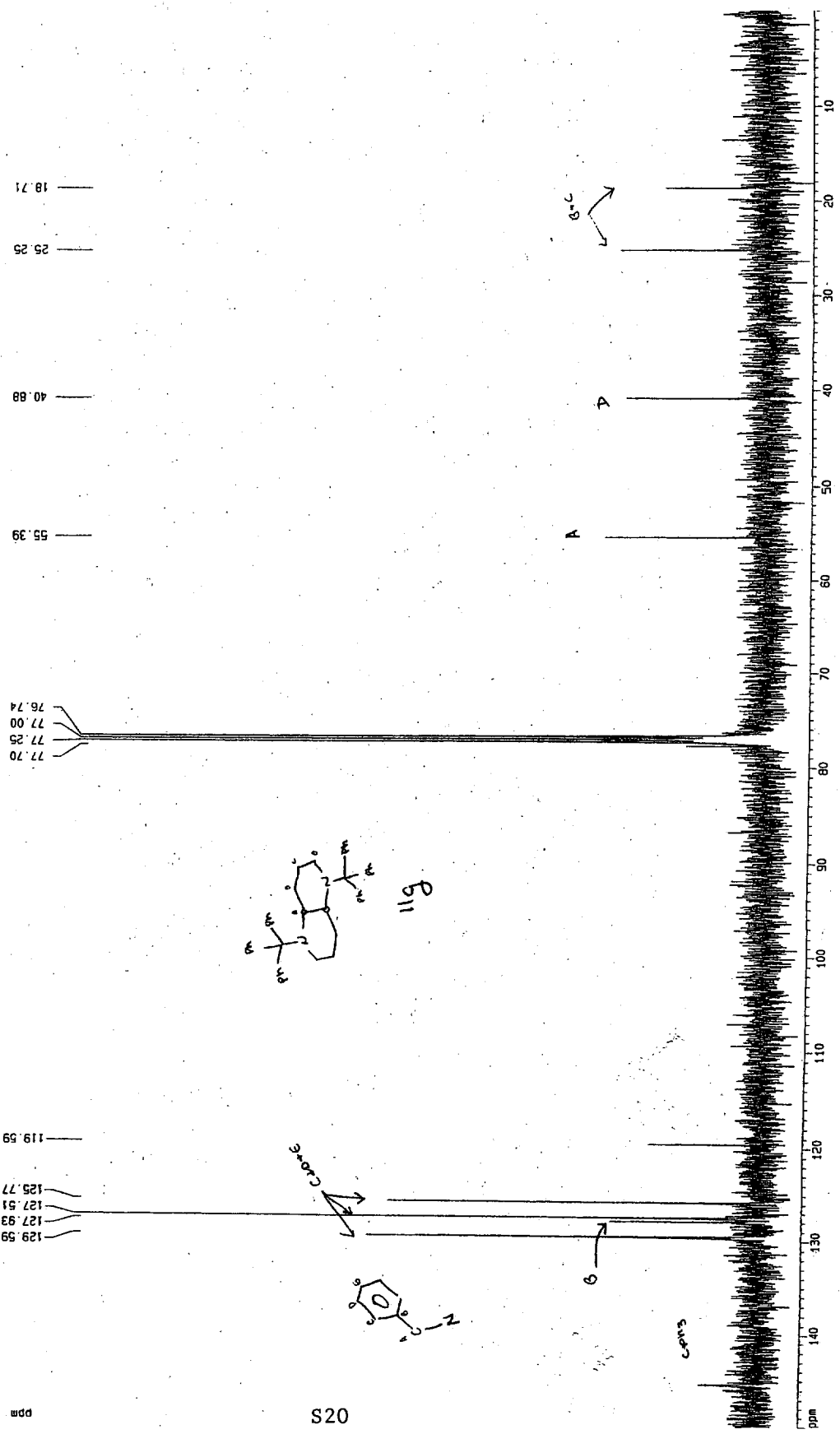


Current Data Parameters  
 NAME 11-174ura  
 EXPNO 1  
 PROCNO 1

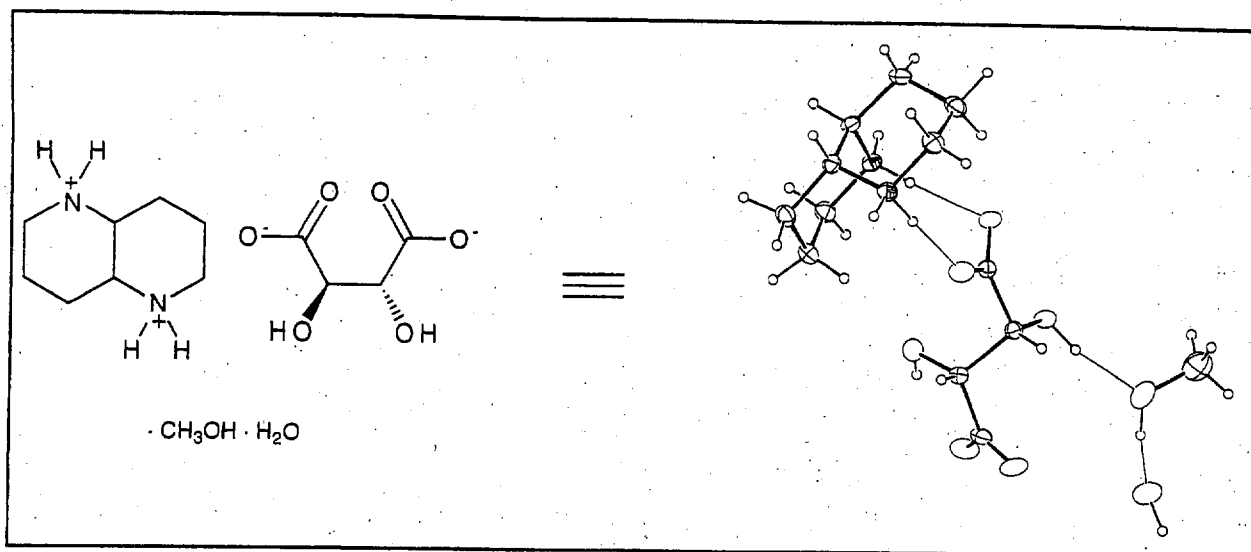
F2 - Acquisition Parameters  
 Date\_ 991012  
 Time 5:14  
 INSTRUM spect  
 PROBD 5 mm Dm1 L1  
 PULPROG zgpg30  
 TD 65536  
 ID SOLVENT CDCl3  
 NS 2  
 DS 2  
 FWH 1620.232 Hz  
 SFO 101.253 MHz  
 AQ 0.683000 sec  
 RG 16384  
 DC 15.000 usec  
 DE 18.75 usec  
 TE 300.0 K  
 H1 40 dB  
 H11 0.0300000 sec  
 S1 11.11111111  
 D1 2.0000000 sec  
 CHOPRG waltz16  
 P31 107.00 usec  
 C2 18 dB  
 P1 5.00 usec  
 DE 18.75 usec  
 SFO1 125.83214 MHz  
 NUC1E13  
 NUC1E13

F2 - Processing parameters  
 SI 32768  
 SF 125.8231675 MHz  
 AQ 0.683000 sec  
 F1 16873.47 Hz  
 F2 0.000000 Hz  
 PRACH 4.28271 dBm/c  
 N2CH 539.24213 Hz/c

1D NMR 0101 operators  
 CL 30.00 cm  
 RP 100.000000 Hz  
 F1 16873.47 Hz  
 F2 0.000000 Hz  
 PRACH 4.28271 dBm/c  
 N2CH 539.24213 Hz/c



## X-ray Structure Determination of Compound 812



Compound 812,  $C_{12}H_{20}N_2O_6$ , crystallizes in the orthorhombic space group  $P2_12_12_1$  (systematic absences  $h00: h=\text{odd}$ ,  $0k0: k=\text{odd}$ , and  $00l: l=\text{odd}$ ) with  $a=15.7034(2)\text{\AA}$ ,  $b=15.7609\text{\AA}$ ,  $c=6.7080(1)\text{\AA}$ ,  $V=1660.23(3)\text{\AA}^3$ ,  $Z=4$  and  $d_{\text{calc}}=1.153\text{ g/cm}^3$ . X-ray intensity data were collected on a Rigaku R-Axis IIc area detector employing graphite-monochromated Mo- $K_{\alpha}$  radiation ( $\lambda=0.71069\text{ \AA}$ ) at a temperature of  $210^\circ\text{K}$ . Indexing was performed from a series of  $1^\circ$  oscillation images with exposures of 200 seconds per frame. A hemisphere of data was collected using  $6^\circ$  oscillation angles with exposures of 500 seconds per frame and a crystal-to-detector distance of 82 mm. Oscillation images were processed using biotex<sup>1</sup>, producing a listing of unaveraged  $F^2$  and  $\sigma(F^2)$  values which were then passed to the teXsan<sup>2</sup> program package for further processing and structure solution on a Silicon Graphics Indigo R4000 computer. A total of 10693 reflections were measured over the ranges  $5.16 \leq 2\theta \leq 50.7^\circ$ ,  $-18 \leq h \leq 18$ ,  $-18 \leq k \leq 17$ ,  $-8 \leq l \leq 8$  yielding 3008 unique reflections ( $R_{\text{int}}=0.0299$ ). The intensity data were corrected for Lorentz and polarization effects but not for absorption.

The structure was solved by direct methods (SIR92<sup>3</sup>). Refinement was by full-matrix least squares based on  $F^2$  using SHELXL-93<sup>4</sup>. All reflections were used during refinement ( $F^2$ 's that were experimentally negative were replaced by  $F^2 = 0$ ). The weighting scheme used was  $w=1/[\sigma^2(F_o^2) + 0.0561P^2 + 0.5062P]$  where  $P = (F_o^2 + 2F_c^2)/3$ . Non-hydrogen atoms were refined anisotropically and

hydrogen atoms were refined isotropically, except methyl hydrogen atoms which were refined using a "riding" model. Refinement converged to  $R_1=0.0406$  and  $wR_2=0.0998$  for 2903 reflections for which  $F > 4\sigma(F)$  and  $R_1=0.0427$ ,  $wR_2=0.1021$  and  $GOF = 1.082$  for all 3008 unique, non-zero reflections and 309 variables<sup>5</sup>. The maximum  $\Delta/\sigma$  in the final cycle of least squares was 0.107 and the two most prominent peaks in the final difference Fourier were +0.194 and -0.302 e/Å<sup>3</sup>.

Table 1. lists cell information, data collection parameters, and refinement data. Final positional and equivalent isotropic thermal parameters are given in Table 2. Anisotropic thermal parameters are in Table 3. Tables 4. and 5. list bond distances and bond angles. Figure 1. is an ORTEP<sup>6</sup> representation of the molecule with 30% probability thermal ellipsoids displayed.

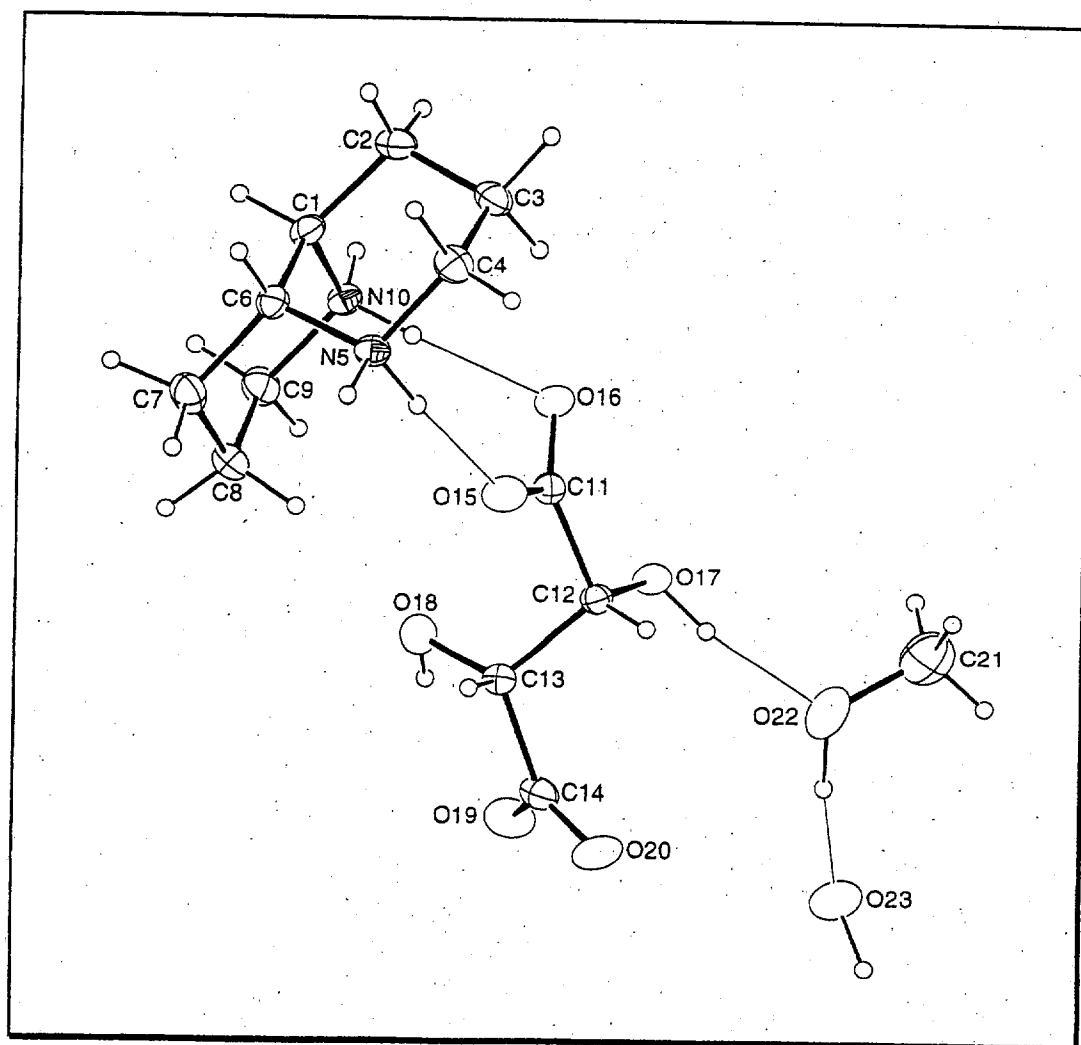


Figure 1. ORTEP drawing of the asymmetric unit with 30% probability thermal ellipsoids.

### References

1. bioteX: A suite of Programs for the Collection, Reduction and Interpretation of Imaging Plate Data, Molecular Structure Corporation (1995).
2. teXsan: Crystal Structure Analysis Package, Molecular Structure Corporation (1985 & 1992).
3. SIR92: Altomare, A., Burla, M.C., Camalli, M., Cascarano, M., Giacovazzo, C., Guagliardi, A., Polidoro, G. (1994). *J. Appl. Cryst.*, **27**, 435.
4. SHELXL-93: Program for the Refinement of Crystal Structures, Sheldrick, G.M. (1993), University of Göttingen, Germany.
5.  $R_1 = \sum ||F_o| - |F_c|| / \sum |F_o|$   
 $wR_2 = \{ \sum w (F_o^2 - F_c^2)^2 / \sum w(F_o^2)^2 \}^{1/2}$   
 $GOF = \{ \sum w (F_o^2 - F_c^2)^2 / (n - p) \}^{1/2}$   
where n = the number of reflections and p = the number of parameters refined.
6. "ORTEP-II: A Fortran Thermal Ellipsoid Plot Program for Crystal Structure Illustrations". C.K. Johnson (1976) ORNL-5138.

Table 1. Summary of Structure Determination of Compound 812

Formula:	$C_{12}H_{20}N_2O_6$
Formula weight:	288.30
Crystal class:	orthorhombic
Space group:	$P2_12_1$ (#19)
Z	4
Cell constants:	
a	15.7034(2)Å
b	15.7609Å
c	6.7080(1)Å
V	1660.23(3)Å <sup>3</sup>
$\mu$	0.93 cm <sup>-1</sup>
crystal size, mm	0.46 x 0.10 x 0.07
$D_{calc}$	1.153 g/cm <sup>3</sup>
F(000)	616
Radiation:	Mo-K $\alpha$ ( $\lambda=0.71069$ Å)
2 $\theta$ range	5.16 – 50.7 °
hkl collected:	-18 ≤ h ≤ 18; -18 ≤ k ≤ 17; -8 ≤ l ≤ 8
No. reflections measured:	10693
No. unique reflections:	3008 ( $R_{int}=0.0299$ )
No. observed reflections	2903 ( $F>4\sigma$ )
No. reflections used in refinement	3008
No. parameters	309
R indices ( $F>4\sigma$ )	$R_1=0.0406$ $wR_2=0.0998$
R indices (all data)	$R_1=0.0427$ $wR_2=0.1021$
GOF:	1.082
Final Difference Peaks, e/Å <sup>3</sup>	+0.194, -0.302



Table 2. Refined Positional Parameters for Compound 812

Atom	x	y	z	U <sub>eq</sub> , Å <sup>2</sup>
C1	0.21644(13)	0.76009(14)	0.4666(3)	0.0280(4)
C2	0.14347(14)	0.6988(2)	0.5004(4)	0.0343(5)
C3	0.17370(14)	0.6066(2)	0.5068(4)	0.0363(5)
C4	0.2247(2)	0.5847(2)	0.3228(4)	0.0371(5)
C6	0.27221(13)	0.73454(13)	0.2909(3)	0.0287(4)
C7	0.3497(2)	0.7920(2)	0.2657(4)	0.0373(5)
C8	0.3996(2)	0.8011(2)	0.4604(4)	0.0367(5)
C9	0.3426(2)	0.82927(14)	0.6296(4)	0.0360(5)
N5	0.29889(11)	0.64374(12)	0.3057(3)	0.0288(4)
N10	0.26820(11)	0.77026(11)	0.6533(3)	0.0276(4)
C11	0.39140(12)	0.60545(12)	0.8007(3)	0.0272(4)
C12	0.47133(12)	0.58418(13)	0.9214(3)	0.0256(4)
C13	0.54226(12)	0.64431(13)	0.8545(3)	0.0276(4)
C14	0.62473(12)	0.6273(2)	0.9672(3)	0.0318(5)
O15	0.39749(10)	0.59416(11)	0.6158(2)	0.0397(4)
O16	0.32711(9)	0.63424(10)	0.8883(2)	0.0329(3)
O17	0.45360(9)	0.59299(11)	1.1265(2)	0.0345(4)
O18	0.51589(11)	0.72988(9)	0.8806(3)	0.0389(4)
O19	0.64775(11)	0.68406(12)	1.0859(3)	0.0486(5)
O20	0.66272(10)	0.55894(12)	0.9325(3)	0.0468(5)
C21	0.4909(2)	0.4293(2)	1.4447(6)	0.0679(9)
H21a	0.5258(3)	0.4001(13)	1.540(3)	0.102
H21b	0.4609(12)	0.3886(11)	1.3646(7)	0.102
H21c	0.4507(11)	0.4646(3)	1.513(3)	0.102
O22	0.54053(14)	0.47777(13)	1.3266(4)	0.0647(6)
O23	0.68259(13)	0.5087(2)	1.5381(4)	0.0523(5)
H1	0.196(2)	0.820(2)	0.449(4)	0.043(7)
H2b	0.116(2)	0.712(2)	0.620(5)	0.051(8)
H2a	0.106(2)	0.701(2)	0.398(4)	0.038(7)
H3a	0.209(2)	0.596(2)	0.639(4)	0.042(7)
H3b	0.122(2)	0.564(2)	0.517(5)	0.069(9)
H4b	0.186(2)	0.590(2)	0.191(4)	0.044(7)
H4a	0.247(2)	0.530(2)	0.329(5)	0.050(8)
H5a	0.337(2)	0.630(2)	0.425(5)	0.054(8)
H5b	0.333(2)	0.630(2)	0.193(5)	0.052(8)
H6	0.235(2)	0.741(2)	0.185(4)	0.039(7)
H7b	0.332(2)	0.851(2)	0.217(5)	0.055(8)
H7a	0.390(2)	0.770(2)	0.157(5)	0.044(7)
H8a	0.426(2)	0.747(2)	0.491(4)	0.034(6)
H8b	0.440(2)	0.846(2)	0.433(5)	0.063(9)
H9b	0.317(2)	0.888(2)	0.614(5)	0.044(7)
H9a	0.372(2)	0.828(2)	0.764(4)	0.039(7)
H10a	0.289(2)	0.719(2)	0.704(4)	0.045(7)
H10b	0.231(2)	0.792(2)	0.748(4)	0.042(7)
H12	0.4890(14)	0.5261(14)	0.885(4)	0.026(5)
H13	0.550(2)	0.634(2)	0.718(4)	0.031(6)
H17	0.489(2)	0.560(2)	1.203(6)	0.065(10)
H18	0.551(2)	0.746(2)	0.976(6)	0.074(11)

H22	0.599(3)	0.498(2)	1.415(6)	0.087(12)
H23b	0.683(2)	0.522(2)	1.652(6)	0.057(10)
H23a	0.731(3)	0.473(3)	1.524(7)	0.11(2)
$U_{eq} = 1/3[U_{11}(aa^*)^2 + U_{22}(bb^*)^2 + U_{33}(cc^*)^2 + 2U_{12}aa^*bb^*\cos\gamma + 2U_{13}aa^*cc^*\cos\beta + 2U_{23}bb^*cc^*\cos\alpha]$				

Table 3. Refined Thermal Parameters (U's) for Compound 812

Atom	$U_{11}$	$U_{22}$	$U_{33}$	$U_{23}$	$U_{13}$	$U_{12}$
C1	0.0242(9)	0.0332(10)	0.0266(10)	0.0026(8)	-0.0033(8)	0.0064(8)
C2	0.0210(10)	0.0468(13)	0.0349(12)	-0.0010(10)	-0.0021(10)	0.0003(9)
C3	0.0280(10)	0.0414(12)	0.0394(13)	0.0013(10)	0.0001(10)	-0.0085(9)
C4	0.0369(12)	0.0356(12)	0.0387(13)	-0.0052(10)	-0.0043(10)	-0.0072(10)
C6	0.0285(11)	0.0319(11)	0.0256(10)	0.0029(8)	-0.0014(9)	0.0011(8)
C7	0.0383(12)	0.0392(12)	0.0344(12)	0.0046(10)	0.0078(10)	-0.0033(10)
C8	0.0290(10)	0.0360(12)	0.0452(14)	0.0041(10)	0.0019(10)	-0.0076(10)
C9	0.0349(12)	0.0340(11)	0.0392(13)	-0.0011(10)	-0.0030(10)	-0.0060(9)
N5	0.0260(9)	0.0324(9)	0.0279(9)	-0.0030(8)	0.0017(8)	0.0012(7)
N10	0.0260(9)	0.0293(9)	0.0275(9)	-0.0020(7)	0.0000(7)	0.0045(7)
C11	0.0234(10)	0.0291(10)	0.0290(11)	0.0033(8)	-0.0028(8)	-0.0021(8)
C12	0.0234(9)	0.0306(10)	0.0229(10)	0.0025(8)	0.0010(7)	0.0015(8)
C13	0.0246(10)	0.0295(10)	0.0287(11)	-0.0004(9)	0.0006(8)	0.0004(8)
C14	0.0223(9)	0.0455(12)	0.0275(11)	0.0007(9)	-0.0002(8)	-0.0054(9)
O15	0.0321(8)	0.0603(10)	0.0266(8)	-0.0016(8)	-0.0041(6)	0.0035(7)
O16	0.0241(7)	0.0420(8)	0.0328(8)	0.0000(7)	-0.0021(6)	0.0048(6)
O17	0.0254(7)	0.0545(9)	0.0237(7)	0.0085(7)	0.0023(6)	0.0064(7)
O18	0.0396(8)	0.0289(8)	0.0481(10)	0.0008(8)	-0.0003(8)	-0.0019(7)
O19	0.0381(9)	0.0626(11)	0.0452(10)	-0.0125(9)	-0.0095(8)	-0.0102(8)
O20	0.0313(8)	0.0603(11)	0.0488(11)	-0.0048(9)	-0.0065(8)	0.0131(8)
C21	0.075(2)	0.066(2)	0.062(2)	-0.006(2)	0.002(2)	0.016(2)
O22	0.0695(14)	0.0508(11)	0.074(2)	0.0161(11)	-0.0261(12)	0.0043(10)
O23	0.0427(10)	0.0682(13)	0.0460(12)	-0.0036(10)	0.0001(9)	0.0162(9)

The form of the anisotropic displacement parameter is:  
 $\exp[-2\pi^2(a^*U_{11}h^2 + b^*U_{22}k^2 + c^*U_{33}l^2 + 2b^*c^*U_{23}kl + 2a^*c^*U_{13}hl + 2a^*b^*U_{12}hk)]$ .

Table 4. Bond Distances in Compound 812, Å

C1-N10	1.502(3)	C1-C2	1.516(3)	C1-C6	1.522(3)
C1-H1	1.00(3)	C2-C3	1.529(3)	C2-H2b	0.94(3)
C2-H2a	0.91(3)	C3-C4	1.511(3)	C3-H3a	1.06(3)
C3-H3b	1.06(3)	C4-N5	1.495(3)	C4-H4b	1.08(3)
C4-H4a	0.94(3)	C6-N5	1.494(3)	C6-C7	1.527(3)
C6-H6	0.92(3)	C7-C8	1.529(3)	C7-H7b	1.02(3)
C7-H7a	1.02(3)	C8-C9	1.511(3)	C8-H8a	0.97(3)
C8-H8b	0.97(3)	C9-N10	1.502(3)	C9-H9b	1.01(3)
C9-H9a	1.01(3)	N5-H5a	1.02(3)	N5-H5b	0.95(3)
N10-H10a	0.93(3)	N10-H10b	0.92(3)	C11-O16	1.253(3)
C11-O15	1.256(3)	C11-C12	1.531(3)	C12-O17	1.411(3)
C12-C13	1.530(3)	C12-H12	0.99(2)	C13-O18	1.422(2)
C13-C14	1.523(3)	C13-H13	0.93(3)	C14-O19	1.251(3)
C14-O20	1.253(3)	O17-H17	0.92(4)	O18-H18	0.88(4)
C21-O22	1.348(4)	C21-H21a	0.96	C21-H21b	0.96
C21-H21c	0.96	O22-H22	1.13(4)	O23-H23b	0.80(4)
O23-H23a	0.95(5)				

Table 5. Bond Angles in Compound 812, °

N10-C1-C2	110.6(2)	N10-C1-C6	111.2(2)	C2-C1-C6	112.5(2)
N10-C1-H1	100(2)	C2-C1-H1	112(2)	C6-C1-H1	110(2)
C1-C2-C3	112.0(2)	C1-C2-H2b	110(2)	C3-C2-H2b	109(2)
C1-C2-H2a	111(2)	C3-C2-H2a	105(2)	H2b-C2-H2a	110(2)
C4-C3-C2	111.0(2)	C4-C3-H3a	111.5(14)	C2-C3-H3a	109.7(14)
C4-C3-H3b	108(2)	C2-C3-H3b	112(2)	H3a-C3-H3b	105(2)
N5-C4-C3	109.5(2)	N5-C4-H4b	109.4(14)	C3-C4-H4b	110.9(14)
N5-C4-H4a	107(2)	C3-C4-H4a	112(2)	H4b-C4-H4a	108(2)
N5-C6-C1	111.3(2)	N5-C6-C7	110.6(2)	C1-C6-C7	112.8(2)
N5-C6-H6	110(2)	C1-C6-H6	102(2)	C7-C6-H6	110(2)
C6-C7-C8	111.7(2)	C6-C7-H7b	111(2)	C8-C7-H7b	109(2)
C6-C7-H7a	112(2)	C8-C7-H7a	109(2)	H7b-C7-H7a	104(2)
C9-C8-C7	111.5(2)	C9-C8-H8a	110(2)	C7-C8-H8a	108(2)
C9-C8-H8b	108(2)	C7-C8-H8b	104(2)	H8a-C8-H8b	114(2)
N10-C9-C8	111.0(2)	N10-C9-H9b	105.3(14)	C8-C9-H9b	115(2)
N10-C9-H9a	104(2)	C8-C9-H9a	114(2)	H9b-C9-H9a	106(2)
C6-N5-C4	112.5(2)	C6-N5-H5a	115(2)	C4-N5-H5a	105(2)
C6-N5-H5b	109(2)	C4-N5-H5b	111(2)	H5a-N5-H5b	104(2)
C1-N10-C9	113.5(2)	C1-N10-H10a	113(2)	C9-N10-H10a	108(2)
C1-N10-H10b	106(2)	C9-N10-H10b	110(2)	H10a-N10-H10b	107(2)
O16-C11-O15	125.1(2)	O16-C11-C12	119.5(2)	O15-C11-C12	115.4(2)
O17-C12-C13	111.7(2)	O17-C12-C11	109.4(2)	C13-C12-C11	107.8(2)
O17-C12-H12	112.8(14)	C13-C12-H12	107.3(13)	C11-C12-H12	107.7(14)
O18-C13-C14	110.7(2)	O18-C13-C12	109.8(2)	C14-C13-C12	111.4(2)
O18-C13-H13	109(2)	C14-C13-H13	110(2)	C12-C13-H13	106(2)
O19-C14-O20	126.5(2)	O19-C14-C13	115.8(2)	O20-C14-C13	117.6(2)
C12-O17-H17	112(2)	C13-O18-H18	100(2)	O22-C21-H21a	109.5(2)
O22-C21-H21b	109.5(2)	H21a-C21-H21b	109.5	O22-C21-H21c	109.5(2)
H21a-C21-H21c	109.5	H21b-C21-H21c	109.5	C21-O22-H22	108(2)
H23b-O23-H23a	105(4)				